

Opioid Receptor Antagonists and Alcohol Dependence Treatment

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PART I: MECHANISMS OF ACTIONS

One of the features of human and animal behavior is the disposition for behaviours experienced as reinforcing to be more likely to occur in the future. In the case of alcohol consumption, the reinforcing effect can happen for two reasons: a) positive reinforcement: alcohol induces satisfying conditions that stimulate new consumptions; with repeated use of alcohol, drinking and stimuli associated with it gradually become more attractive, leading to a loss of control and creation of an impulse focused on alcohol consumption; b) negative reinforcement: alcohol is consumed to relieve or avoid withdrawal symptoms; the reduction or suspension of the substance gives rise to neurodegenerative and affective disorders, with alcohol consumption arising from the need to suppress such negative changes [1].

Both positive and negative reinforces are condition the desire to repeat the consumption of alcohol through processes mediated by neuronal changes. It's when, how and where such changes occur, that transform the individual into an alcohol dependent person, is the goal of the neurobiological studies on alcohol, aiming that such knowledge may contribute to reduce the consequences of alcohol abuse and foster better therapeutic interventions (Figure 1).

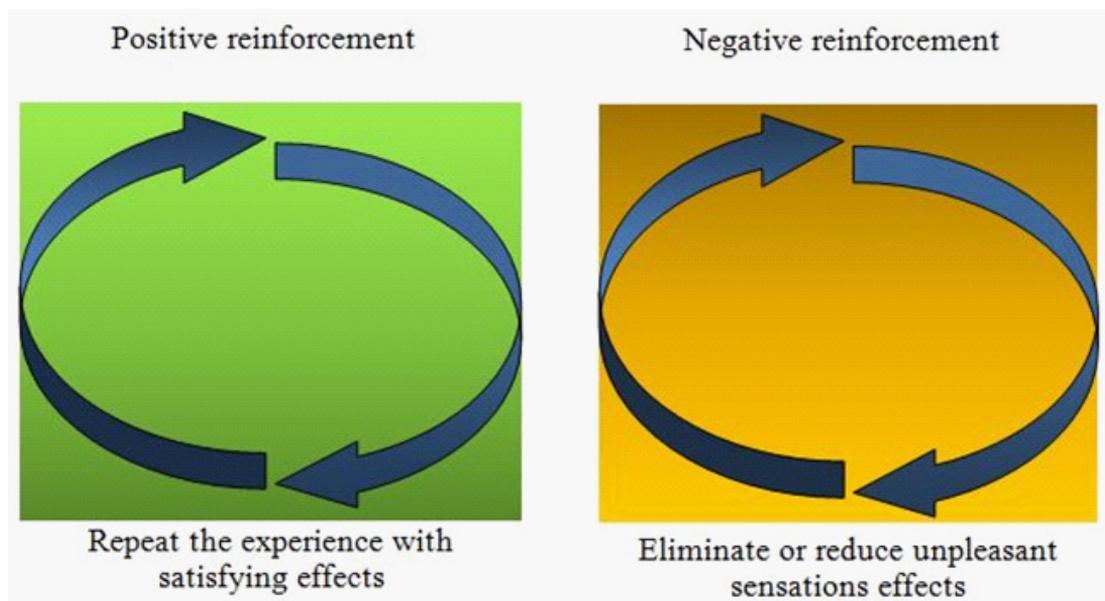


Figure 1: Positive and Negative Reinforcement.

DOPAMINE AND THE BRAIN REWARD SYSTEM

When explaining the satisfying action of alcohol, there is abundant evidence that ethanol promotes activation of the mesolimbic-mesocortical dopaminergic path. These bundles project from the ventral tegmental area (**VTA**) to the dopamine receptors (**DA**) located in the *nucleus accumbens* (**NAC**) and other mesolimbic areas, such as the hypothalamus, amygdala, hippocampus, septal area and pre-frontal cortex [2-5], constituting the central circuit of the brain reward system involved in the regulation of mnemonic processes learning of motivated behaviours, modulation of affection and externalization of emotions by activating motor pathways. Repeated stimulation of dopaminergic neurons by the action of alcohol, associated with the release of DA in the *nucleus accumbens*, emerges as the key mechanism for the onset and maintenance of alcohol consumption preference [2,6].

Numerous animal studies show the increase in extracellular levels of dopamine in the *nucleus accumbens* [2,7,8] and amygdala [9] after injection or self-administration of alcohol, with such increase being higher in alcohol-preferring genetically selected animals, which indicates the influence of genetic factors in the activation of dopamine and, ultimately, in the reinforcing properties of alcohol [7,10].

Genetic polymorphism has also been evidenced in clinical and laboratorial studies in humans, highlighting the participation of allele variants of the D2 and D4 receptor genes in vulnerability to alcohol dependence [11-14]. On the other hand, studies carried out in young people at high risk of alcohol dependence show greater increase in extracellular DA levels than the control groups without this associated risk [15].

Besides the release of DA, alcohol also affects DA receptors, particularly the D2 receptor. Studies using different methodologies found that strains of alcohol-preferring genetically selected animals are distinguished by the reduction of D2 receptors concentration in the limbic system [16], whereas human subjects with alcohol dependence show a lower affinity with D2/D3 receptors than the control groups [17-20], suggesting an increased vulnerability to substance abuse. Observations carried out both in animals and in humans associate high levels of D2 receptors to more intense responses to alcohol intoxication, which seems to constitute a protective factor against substance abuse, whereas subjects with low levels of D2 receptors show a more attenuated response and greater propensity for continued consumption of alcohol and increased risk of alcohol dependence [16-18,20-22]. D1 receptors also seem to be involved in a down regulation system in response to alcohol but results have been less conclusive [23].

DA receptors agonist and antagonist substances have led researches to inconsistent results, showing that both have a reducing action of the reinforcement induced by alcohol, favouring the reduction of consumption. Agonist substances (bromocriptine, lisuride, apomorphine, modafinil) reduce dopamine deficit and reverse the preference for alcohol, but the success observed with animals was not seen in humans [24-27]. In the second case, antagonist agents (haloperidol, tiapride, risperidone, flupentixol) reduce the hyperactivity induced by low doses of alcohol, attenuate the reinforcing effect of drinking and decrease symptoms associated with withdrawal syndrome, an action related to the affinity for D2 receptors [28,29] but evidence of efficacy in maintaining abstinence are scarce [27-30].

In convergence with the positive reinforcement hypothesis, the dopaminergic system is also involved in the negative reinforcement effect of alcohol. Animals with symptoms of alcohol withdrawal present levels of extracellular dopamine below normal in the *nucleus accumbens*; as the reduction of dopamine in this area of the brain may predict depression/dysphoria, this change plays an influential role in the symptomology of alcohol abstinence and negative reinforcement that accompanies this syndrome [31,32]. It was found recently that chronic exposure to alcohol changes the function of D2 and D4 receptors in the pre-frontal cortex, with these changes being maintained during the initial stages of abstinence, which contributes to the deficit of executive functions, such as decision making and cognitive control of behaviour, even after cessation of ethanol consumption [33]. Linking these actions there is a two-phase model that differentiates low consumption situations from abuse/dependence situations [32]. In low consumption situations, alcohol triggers a complex set of stimuli and responses that lead to a feeling of well-being and

ultimately to positive reinforcement. In cases of continued, excessive consumption, disruption of cellular interactions leads to a two-stage process: in the first stage, alcohol causes a brief feeling of well-being associated with the release of the neurotransmitters in the reward circuits, but as ingestion of alcohol continues, the synthesis and release of neurotransmitters changes, causing deficit of brain messengers, along with anxiety, irritation, or craving, and leading to a repeated self-sustained search pattern for alcohol [32].

Several mechanisms have been proposed to explain the influence of ethanol over dopaminergic activity but the process remains under discussion [4,6,34-36]. Regardless of the mechanism involved, some researchers consider the “dopaminergic hypothesis” as the neurochemical explanation that best explains the development of motivation and dependence from alcohol [1,3] whereas others consider that the simple activation of the dopaminergic system is insufficient for understanding the mechanisms underlying the feeling of pleasure and reinforcement induced by alcohol and even claim not to be necessary the integrity of mesolimbic dopaminergic neurons to cause the alcohol dependence process [35]. Currently, data point to the existence of neuro circuits through which the endogenous opioid system acts in the release of DA in the *nucleus accumbens* and the mechanisms responsible for the ethanol reinforcing effects [34,37-40].

ENDOGENOUS OPIOID SYSTEM

Endogenous opioid peptides are non-classical neurotransmitters with an activity similar to that of opioid analgesics. Based on the pharmacological profile, three different subfamilies were initially identified and characterized – endorphins, enkephalins and dynorphins – and three types of inhibitory G protein-coupled receptors, designated as *mu* (μ), *kappa* (κ) and *delta* (δ) (respectively MOP, KOP and DOP, using the terminology recommended by IUPHAR - International Union of Pharmacology) [41]. Later on, new families were isolated, named nociceptin and endomorphins, but their functions and clinical relevance are still under study [42-44].

Endogenous opioid peptides originate from inactive precursor molecules (pro-opiomelanocortin - **POMC**; pro-enkephalin; pro-dynorphin), with similarities in the organization of their genes, which suggests a common ancestor [42]. The processes by which precursor molecules form active peptides are not equal in all tissues and the same precursor may lead to different products depending on the tissue and the receive signal. Through a set of tissue-specific enzymatic stages, the precursor protein pro-opiomelanocortin originates the melanocyte-stimulating hormones (**MSHs**), corticotrophin (**ACTH**) β -lipotropic pituitary hormone (**β -LPH**) and four types of endorphins (α , β , γ , δ) including β -endorphin, the most powerful endogenous opioid peptide neurotransmitters [45]. Pro-enkephalin originates two forms of enkephalin, the methionine-enkephalin (met-enkephalin) and the leucine-enkephalin (leu-enkephalin). Pro-dynorphin encodes dynorphins, chains of aminoacids with different lengths, with the leu-enkephalin sequence: dynorphinA, dynorphin B and α/β -neo-endorphin [36,42,45].

The opioid compounds and their receptors form the endogenous opioid system, involved in a large group of functions encompassing the control of feelings, emotions and affection: modulation of response to painful and stressing stimuli; reward and reinforcement; homeostatic adaptive functions, such as body temperature regulation, ingestion of food and water, reproduction [36,42].

Both the endogenous opioid peptides and the opioid receptors are widely distributed by the central nervous system (CNS) and peripheral nervous system (PNS) mediating the neuromodulator or neurotransmitter functions [37-39]. The β -endorphin is produced by the anterior pituitary, being released into the peripheral systemic circulation, and by the hypothalamic POMC neurons and released in the CNS. The β -endorphin produced in neurons is present in the ventromedial arcuate hypothalamic nucleus and projected to other areas of the hypothalamus (supraoptic nucleus, paraventricular nucleus, and lateral hypothalamus) as well as to the amygdala, ventral tegmental area (VTA), periaqueductal gray and bed nucleus of the *stria terminalis* [46,47]. It is released during pain and stress situations, causing an analgesic effect and reducing the anxiety and causing feelings of euphoria [45].

Enkephalins are also released by neurons in the central nervous system and by cells in the adrenal medulla; in the CNS, they can be found in most brain regions including most hypothalamic and thalamic nucleus, preoptic area, septum, *nucleus accumbens*, ventral tegmental area, amygdala and neocortex, participating in a large number of effects including perception of pain, regulation of memory and emotional conditions, ingestion of food and liquids, immune responses and control of the gonadal function [46]. Dynorphins have been identified in many different parts of the brain (hippocampus, amygdala, hypothalamus, striatum, spinal cord) and intestine and play a regulatory role in numerous functions involving analgesia, reinforcement, stress response, cognitive function (learning and memory) and motor integration [45,48].

Once synthesised and stored in neuronal vesicles, opioid peptides are released in response to specific stimuli that cause depolarization of the neuronal membrane. Opioid peptides disseminate into the synaptic area to interact with the various classes of opioid receptors present on both pre and post-synaptic membranes of opioid and opioid target neurons, located close to the release area (enkephalines) or more distant areas (β -endorphines, dynorphines) [39], being rapidly deactivated at extracellular level by the peptidase enzymes which breakdown active peptides to produce inactive metabolites. Blocking the inactivation of opioid peptides increases their basal extracellular levels near the release site [43,49].

The relationship between endogenous opioid peptides and their receptors is complex. β -endorphins bind with about equal affinity to μ and δ -opioid receptors, whereas enkephalins bind with much greater affinity to δ -opioid receptors than to μ -opioid receptors, and dynorphins tend to bind selectively to κ -opioid receptors [36,37,42,50]. Opioid receptors have various subtypes (μ_1 , μ_2 ; δ_1 , δ_2 ; κ_1a , κ_1b , κ_3 , κ_4) [51] so the results of such interactions must be carefully assessed, avoiding any generalization; however activation of μ - and δ -opioid receptors frequently

leads to euphoria patterns whereas stimulation of k-opioid receptors is associated with dysphoria [37,50,52]. For instance, in the mesolimbic system, β -endorphins and enkephalins mediate the increase of DA release in the *nucleus accumbens* through interaction with μ - and δ -opioid receptors, actively participating in the reward and reinforcement processes [37,38,53]. Dynorphins, on the other hand, lead to inhibition of DA release as a result of activation of k-opioid receptors, creating aversive states [53]. Although it seems that the various sub-types of opioid receptors have a relatively equivalent distribution in rodents brains and in the human brain, differences have been reported that must be considered when using animal models; these differences include a larger k receptor mRNA expression in different areas of the brain, k and μ receptors in the cortex and hippocampus and μ -opioid receptors in the hypothalamus of humans, compared to rats [37,52].

ETHANOL AND ENDOGENOUS OPIOID SYSTEM INTERACTIONS

Although there is a very considerable number of researches that show changes in endogenous opioid peptides as a result of alcohol ingestion, the mechanism of interaction of these molecules in alcohol dependence is still not sufficiently clarified [36-38,40,54,55].

Acute administration of ethanol stimulates the dose-dependent release of endorphins and enkephalins in the hypothalamus and pituitary gland of rodents [36,39,56]. But, although studies have been relatively conclusive on endorphins, the results of ethanol action on the release of enkephalins are contradictory [37,40,50]. Less information is available on dynorphins but studies point to an increased release in the *nucleus accumbens* with exposure to very high doses of alcohol, which may be associated to an aversive effect [48,50,57].

As to extended exposure to alcohol, there is generally a decrease in the positive reinforcement properties from the endogenous opioid system [37,51]. Chronic administration of alcohol decreases POMC gene expression, β -endorphins release and μ -opioid receptors affinity [58], while increasing pro-dynorphins and dynorphins involved in the negative reinforcement effect of alcohol [59-61].

Assessment of ethanol effect over opioid receptors proves to be more difficult, with results depending on the studied areas of the brain, the experimental conditions of alcohol administration and the species and strain of animals studied [36,37]; consequently, studies are frequently inconsistent [36].

An important source of data on the involvement of the endogenous opioid system in alcohol dependence derives from studies in animals and humans with a genetic predisposition for alcohol consumption.

Rats belonging to selected strains due to their high preference for self-administration of alcohol show lower levels of endogenous opioids before alcohol consumption and, compared to animals without such preference (**NP**), the ingestion of ethanol causes an increased release of hypothalamic β -endorphins [36,56,62] and a higher genetic expression of the hypothalamic

[56,62-64] and pituitary gland [65] mRNA POMC. In what concerns enkephalin levels, there are also differences in P and NP strains, with the former showing higher levels of enkephalin in the *nucleus accumbens*, ventral tegmental area [60] and hypothalamus [66] and higher pro-enkephalin mRNA expression in the pre-frontal cortex [63]. In a study where met-enkephalin levels were similar in the striatum, hypothalamus, medulla and pons, the strain with higher pre-disposition for voluntary ethanol consumption had lower pro-enkephalin concentration and met-enkephalin in the mid brain and exhibited marked mesolimbic enkephalin increase following voluntary ethanol consumption [67]; In the same sense, in another study, both strains of animals showed equal baselines of pre-proenkephalin mRNA (**PPENK mRNA**); after infusion of alcohol rats selected due to preference for alcohol showed a significant increase in PPENK mRNA in the *nucleus accumbens* [68]. With dynorphins, results are different, showing a higher number of binding areas and affinity for these peptides and for pro-dynorphin mRNA in the *nucleus accumbens* and septum of ethanol-avoiding mice [69].

Likewise, the alcohol-preferring strains vs alcohol-aversive strains have showed differences in the distribution of opioid receptors. The alcohol-preferring strains tend to show higher density of μ -opioid receptors in the *nucleus accumbens* area and other limbic system structures and in the pre-frontal cortex area [63,70], higher density of δ -opioid receptors in the ventral tegmental area and *nucleus* [71] and lower density of κ -opioid receptors in the ventromedial hypothalamus [63] and *nucleus accumbens* [69] than alcohol-aversive rats. In the past decades, studies performed with genetically manipulated animals showed that animals without the μ -opioid receptor (knockout μ) present a reduction in voluntary alcohol consumptions whereas animals without the δ -opioid receptor (knockout δ) present an increase [72].

Although not being conclusive as to a causal relation, these results suggest a relation between the opioid system and the self-administration of ethanol, highlighting the fact that, in general, these studies show that alcohol-preferring strains present low levels of β -endorphin and enkephalin activity in several areas of the brain, at baseline, and a significant increase as response to acute ethanol exposure [36,56,73].

Similar results were observed in humans. Non-alcohol dependent adults with alcohol dependent parents (high risk group) show lower baseline levels of β -endorphins than non-alcohol dependent adults without a history of alcohol dependence in the family (low risk group) [73]; with alcohol, adults with alcohol-dependent parents show a marked increase of β -endorphins when compared to adults with non-alcohol dependent parents, tending to equate the final results [73,74] although not all studies confirm this discrepancy [75]. Comparing alcohol dependent and non-dependent adults, the former show lower levels of β -endorphins both in plasma and in the CFS, regardless of the alcoholisation time [76,77] which strengthens the hypothesis of β -endorphins binding with a higher sensitivity to alcohol and to genetic pre-disposition to alcohol dependence. Recent studies using PET in humans showed that alcohol causes the release of endorphins in the *nucleus*

accumbens (**NAc**) and orbitofrontal cortex (**OFC**) both in individuals with high consumption of alcohol and in healthy control individuals, with a greater increase in the first group with a positive correlation between changes in the OFC and the extension of alcohol-related problems and the subjective sensation of intoxication [78].

The complexity of the relation between the endogenous opioid system activity and the development of alcohol dependence originated contradictory theories, such as the opioid deficit theory [79,80] and the opioid surfeit theory [81]. Other models relate the opioid system and the use of alcohol to stress and individual differences in the opioid system to alcohol sensitivity [46,74].

In stress-inducing situations, the hypothalamus-pituitary-adrenal (**HPA**) axis is activated mainly by the production of the corticotropin-releasing hormone (**CRH**) which is transported to the receptors on the anterior pituitary and originates the production of β -endorphins and ACTH from its precursor POMC. ACTH influences the adrenal gland to stimulate the release of glucocorticoids. Between the production of endogenous opioids and the production of ACTH there is a negative feedback effect, i.e., endogenous opioids may inhibit the production of ACTH [82]. Similarly to stress, alcohol also stimulates the synthesis of CRH in the hypothalamus and activation of the HPA axis [76,83] with subsequent synthesis and release of endogenous opioids thus contributing to the alcohol reinforcement properties.

Throughout researches, evidence has been stacking on the relation between alcohol consumption and changes in the HPA axis function and the activity of the endogenous opioid system, both in animals [55] and in humans [74,84,85]. Among other data these studies suggest that: reduced β -endorphin baselines and exaggerated responses to stress induce a higher vulnerability to ethanol abuse [86]; the chronic deficiency of β -endorphin is associated with states of higher anxiety and increase of sensitivity to ethanol rewarding properties, at least partially, as it relieves anxiety [55]; reduction of β -endorphin and ACTH activity with chronic consumption of alcohol [84] and deregulation of the HPA axis in withdrawal syndrome [87]; reduction of ACTH response or cortisol response to stress predicts an earlier return to drinking [87]; β -endorphin plasma levels are inversely related to anxiety self-assessment levels during withdrawal [88]. Non-dependent individuals with a family history of alcohol dependence show higher baseline levels of ACTH and an intense endocrine response to stress, dampened by alcohol administration [85]. In summary, these studies point to the fact that alcohol-induced reduction of increased baseline levels of anxiety are a strong factor for negative reinforcement regarding alcohol dependence maintenance [55,85,88,89].

A relevant set of studies supporting alcohol interaction with the endogenous opioid system derives from the use of pharmacological agents acting over the system receptors.

These studies show that μ - and δ -opioid receptors agonists (e.g.: morphine, DAMGO, DALA, DPDPE) increase the release of DA [38,53] and alcohol consumption in laboratory animals

[90], whereas opioid receptor antagonists (naloxone, naltrexone, nalmefene) compete with the endogenous opioid peptides for binding to the opioid peptide receptor and reduce the alcohol-induced dopaminergic activity [38,91], reduce consumption, both in animal models [90,92] and in humans [93-95], reduce craving [96] and relapses [94,97].

Overall, researchers suggest a strong participation of the endogenous opioid system in the stimulation and reinforcement of alcohol consumption. As such, there is a consensus that comprehension of basic mechanisms favouring onset and maintenance of alcoholic drinks consumption must include the intervention of neuropeptides and opioid receptors. Many of the mechanisms underlying the interaction between the opioid system and other neurotransmitter and neuromodulator systems (serotonergic, GABAergic, noradrenergic, cholinergic, cannabinoid...) [39,40,47] are still unclear, as well as the meaning and significance of such changes in alcohol addictive behavior. Not with standing, researches and empirical studies have already led to a point where two out of the four substances approved for pharmacological treatment of alcohol dependence are directly related to the endogenous opioid system.

PART II: CLINICAL PERSPECTIVE

FROM N-ALLYLNORCODEINE TO NALTREXONE AND NALMEFENE

The History of Opioids receptors antagonists began in 1915 when von Julius Pohl first describes the ability of N-allylnorcodeine to reverse effects of morphine and heroin on respiratory depression [98]. But it was not until the 1950s, that Pohl's works are updated and utilized by Hart to synthesized N- allylnormorphine (Nalorphine), an antidote for morphine with analgesic properties [99-102]. However, because of its dysphoric properties, it was rapidly abandoned. Then surged pentazocine and cyclazocine. Cyclazocine have been extensively studied for its potential in opiate addiction, despite promising results, the induction of sleepiness, drunkenness and in some cases hallucinations and respiratory depression prevented its used in clinical practice [103-108]. Naloxone, an allyl derivative of noroxymorphone and the first opioid antagonist reaching important clinical application, became the treatment of choice of opioid overdose [109]. If on one hand, naloxone was a pure opioid antagonist¹ with fewer adverse effects than its predecessors, on the over hand its poor oral bioavailability and short duration of action limited its use for the treatment of opioids or alcohol dependence [110-112]. In an attempt to avoid these disadvantages, Endo laboratories synthesized in 1963, Endo 1639A, a cyclopropylmethyl analog of naloxone, rename Naltrexone [113,114]. It's a relative pure and potent μ -opioid antagonist with longer duration of action and better bioavailability then naloxone [115]. Rapidly it becomes a treatment of choice for opioid addiction and in 1994 was approved by the American Food and drug Administration for the treatment of alcohol dependence. Finally, in 2012, Nalmefene, an opioid antagonist, structurally and functionally related to Naltrexone was approved by the European Medicines Agency (**EMA**) for the treatment of alcohol dependence (Figure 2).

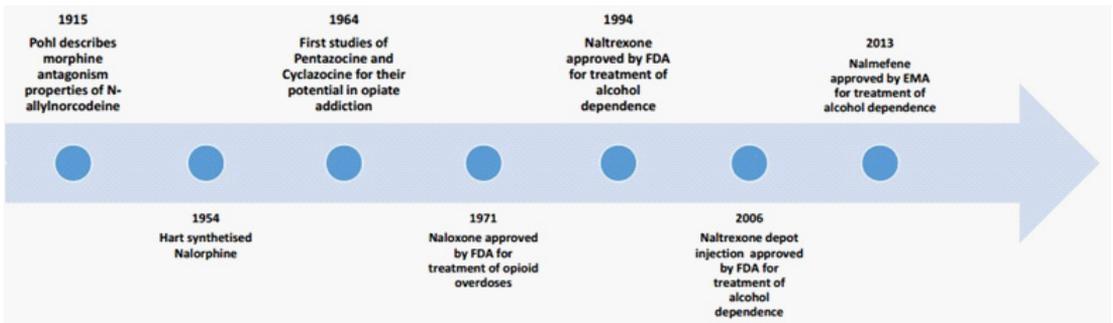


Figure 2: From N-allylnorcodeine to Naltrexone and Nalmefene.

NALOXONE THE PRECURSOR

Naloxone is an allyl derivative of noroxymorphone and a potent non-selective opioid antagonist, with an affinity order of: $\mu > \kappa \gg \delta$ - opioid receptors [110,116] (Figure.3). It was the first opioid antagonists evidencing potential efficacy as a treatment for alcohol dependence in animal experimentation. For example, Naloxone abolishes the preference for alcohol developed by rats after 15 days of administration of an ethanol solution [117]. The injection of Naloxone also decreased alcohol self-administration, in rats educated to press a lever to obtain water or alcohol, [118]. This suppressing effect on alcohol intake, was also observe in rats which were selectively bred alcohol preference [119]. However, despite this potentialities, its short duration of action, reduce bioavailability, the need of very high doses to sustain opiate blockade (2 - 3g for 24h), and the discovery of new opioid antagonists, like naltrexone, prevented its clinical use for the treatment of alcoholism in man [112,120,121].

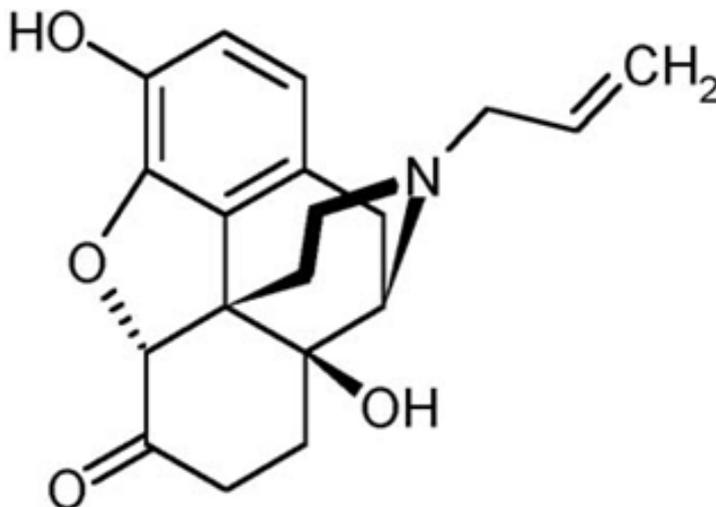


Figure 3: Chemical structure of Naloxone.

NALTREXONE FOR THE TREATMENT OF ALCOHOL DEPENDENCE

Chemistry and Mechanism of Action

Naltrexone is an anicyclopropyl derivative of oxymorphone, structurally similar to naloxone and is also an opioid antagonist with a high affinity for μ -opioid receptors, an intermediate affinity for κ -opioid receptors, and a very low affinity for δ -opioid receptors [116,122,123] (Figure.4). It is believed that naltrexone blocks opioidergic inhibition of GABAergic inhibitory interneurons in the ventral tegmentum area and by so blocks release of dopamine in the nucleus accumbens, neutralizing the increase in dopaminergic neurotransmission produced by alcohol consumption, and ultimately diminishing its pleasurable effects [124]. Orally, it is well and rapidly absorbed, but suffers an extensive first-pass metabolism in the liver, where it is metabolized to 6- β -naltrexol. It is mainly excreted in the urine. The Half-life of Naltrexone ranges between 4 and 10 hours dependent on the duration of the administration, and is of 12 to 16 hours for 6- β -naltrexol [125,126].

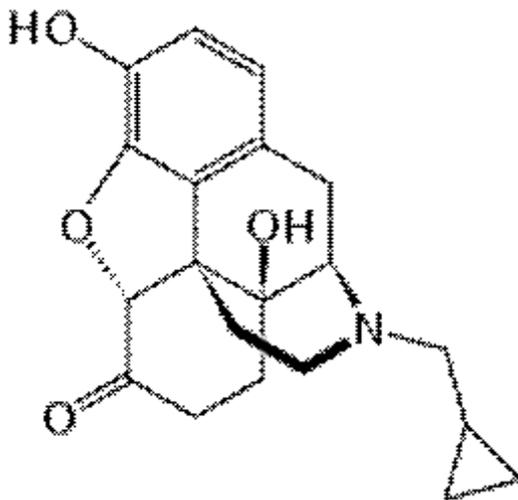


Figure 4: Chemical Structure of Naltrexone.

Efficacy

Naltrexone efficacy in the treatment of alcohol dependence was initially supported by two double blind, placebo-controlled trials of Volpicelli et al and O'Malley et al. They evidenced that 50mg of oral Naltrexone in adjunct with psychosocial alcohol rehabilitation programs (Volpicelli study) or Coping Skills Therapy oriented for abstinence (O'Malley study), significantly decreased alcohol craving, mean of drinking days, and rates of relapse [93,94]. A large multicenter study, the COMBINE study, demonstrated that 100mg of oral Naltrexone associated with medical management is more effective than placebo or Acamprosate in reducing drinking days and heavy drinking [127, 128]. Compared to placebo, Naltrexone reduced percent days of abstinence

in 5,5% ($p=0,02$) and risk of heavy drinking days (hazard ratio, 0.72; $P=.02$) [127]. Many over studies with different endpoints and different adjunctive treatments confirm the effectiveness of Naltrexone in alcohol dependence. However, an important multicenter clinical trial sponsored by the Department of Veterans Affairs which enrolled men with chronic and severe alcohol dependence found no significant differences between Naltrexone and placebo for the prevention of relapse, or for reducing the amount of drinks per day. Finally, a recent Cochrane review based on 50 trials including 7793 patient, concluded that Naltrexone is a safe and effective strategy in alcoholism treatment with a moderate size effect in the reduction of heavy drinking and drinking days [129]. On this review Naltrexone was found to reduced drinking days by 3,9%, and the risk of heavy drinking by 83% [129].

Management

Naltrexone Hydrochloride (REVIA®; DEPADE®), was approved by the US Drug and Food Administration (FDA) in 1994 for the treatment of alcohol dependence as part of an appropriate plan management for the addiction. The treatment should be initiated after at least 3 days of abstinence, and a medical evaluation with physical exam, psychosocial evaluation and laboratory testing including liver enzyme and screening test for drugs of abuse must precede the prescription. Naltrexone should be start at 25mg per day and increase to a maintenance dose of 50mg per day for up to 6 month or longer if the patient wants to continues. However, the drug should be stopped if drinking persist 4 to 6 week after initiating Naltrexone [130-132].

The 2011 NICE guidelines distinguish two situations in which Naltrexone can be used [131]:

- First, after a successful withdrawal, Naltrexone can be considered in combination with a psychological intervention, in moderate and severe alcohol dependence. The aim is to maintain abstinence.
- Secondly, Naltrexone in combination with a psychological intervention can be used for harmful drinkers and people with mild alcohol dependence who has not responded to psychological interventions alone.

The 2009 Treatment Improvement Protocol of the U.S. Center for Substance Abuse Treatment Highlights that oral Naltrexone should be used in patients who are highly motivated or associated to a medication monitoring plan. It is also put forward that Naltrexone in patient with intense craving or with a family history of alcohol dependence may be more beneficial [130,132].

The 2015 French guidelines of the *Société Française d'Alcoologiesubline* that Naltrexone associated with psychosocial intervention is a first line treatment for preventing relapses after alcohol withdrawal [133].

Adverse Effects and Limitations of Use

Some mild and time limited side effect can be experience in up to 30% of patient, including gastrointestinal symptoms, like nausea, vomiting, or abdominal pain, and symptoms associated

with a decreased arousal, like fatigue, sleepiness or weakness. Decreased libido and depression can also be experienced [129]. Naltrexone in High doses (100 to 300mg/day) was associated with hepatotoxicity in studies with obese patient and that why liver enzymes should be monitored and Naltrexone is contraindicated in liver failure [134,135]. Another concern is about patient using illicit opioid drugs, buprenorphine, methadone, opioid analgesic or medications containing an opioid, because of the necessary risk of acute withdrawal syndrome, and is contraindicated in this cases [130,132].

Extended-Released Injectable Naltrexone (XR-NTX)

Although evidencing efficacy, oral Naltrexone have several limitations, and adherence is probably the big one. Specific studies point out the impact of adherence on efficacy, for example a multicenter, randomized, double blind clinical trial comparing Naltrexone and placebo in 175 patient find no differences in drinking outcomes, however when only compliant patient were analyzed, Naltrexone appears to be superior to placebo [136]. Another one comparing Naltrexone and placebo in 97 patients also evidenced comparable results [137]. A reanalyzes of two major clinical trials that failed to encounter efficacy of Naltrexone over the placebo also revealed that medication compliance enhanced the effect of naltrexone [138]. Alongside with these clinical trials, naturalistic data are even more striking. A retrospective analysis of two commercial, community-based, claims database revealed that of 1138 patients who were prescribed oral Naltrexone and filled an initial prescription, 51,8% filled only one prescription and the vast majority (85,8%) didn't persist on Naltrexone (defined as having filled prescriptions for $\geq 80\%$ of the 6-month treatment period) [139]. They also evidenced that non-persistent patient had a poorer outcome with significantly more emergency room admission and hospitalizations. In the same way another retrospective analysis of a claims database evidenced an even poorer treatment adherence [140] (Figure 5).

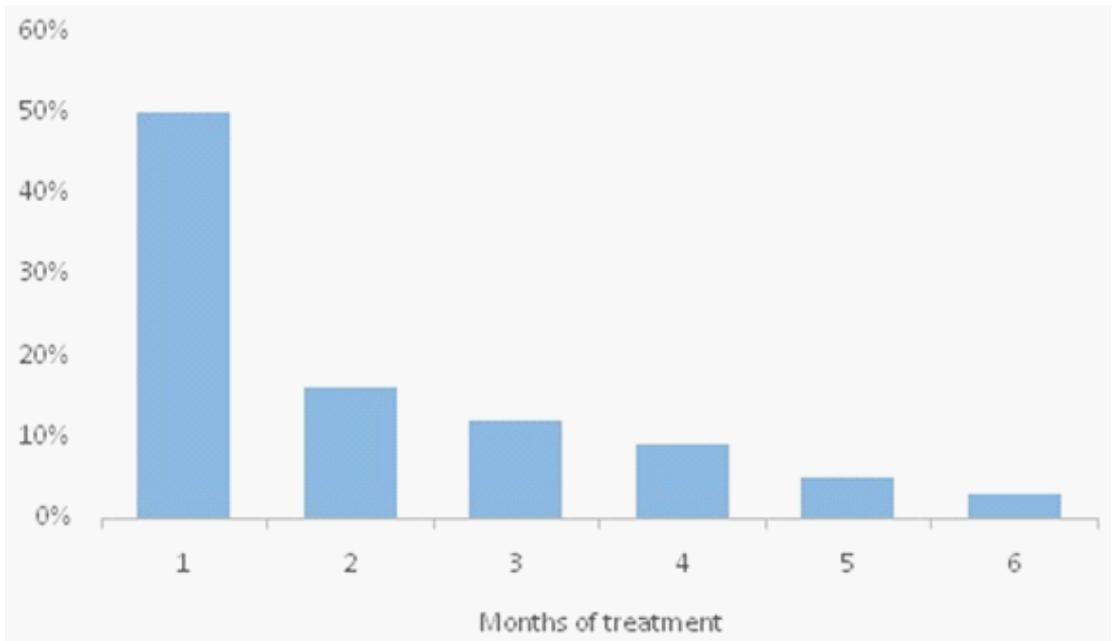


Figure 5: Estimated adherence to oral Naltrexone across time based on literature data.

It is in this context that a Naltrexone sustained-release formulation is meaningful.

In fact, an extended-released injectable suspension of Naltrexone (VIVITROL®) was approved by the U.S. FDA for the treatment of alcohol dependence in 2006. It is a formulation containing microspheres of polylactide-co-glycolide loaded with Naltrexone. An injection contains 380mg of Naltrexone and has to be administered into the gluteal muscle once every 4 weeks. XR-NTX avoid first-pass hepatic metabolism permitting a monthly total doses 4-fold less in comparison with oral Naltrexone. Moreover XR-NTX yield plasma area under the curve 3 to 4 times higher than oral Naltrexone, with more stable levels [141].

The efficacy of XR-NTX was demonstrated in a multicenter randomized controlled trial including 624 participants, receiving a monthly injection with XR-NTX 380mg, XR-NTX 190mg or placebo, combined with a 12 sessions of low-intensity psychosocial intervention. Compared with placebo XR-NTX 380mg decreased in 25% the rate of heavy drinking days [142]. A secondary analysis of outcomes determine that XR-NTX 380mg was more effective in reducing drinking days and heavy drinking days in patient with at least 4 days of abstinence, while XR-NTX show intermediate results indicating a dose-effect response [143]. A Cochrane review indicate that XR-NTX reduced the risk of any drinking after detoxification to 92% of the placebo [129].

XR-NTX have a safety profile similar to oral Naltrexone excepted for injection site reactions, and theoretically lower risk of liver toxicity because of its lack of first-pass hepatic metabolism, and reduced doses [129,142].

The 2009 Treatment Improvement Protocol of the U.S. Center for Substance Abuse Treatment suggest that XR-NTX is probably a better option for patient in which adherence to oral medication is an issue or in those who prefer not have to remember to take daily oral medication [132].

Factors Influencing the Response to Naltrexone

Numerous potentials factors associated with a better response to Naltrexone in alcohol dependence have been suggested like: family history of alcoholism [144-147]; history of abuse of other substances [144]; onset of alcohol abuse before age 25 [144,148]; higher levels of craving [145,149]; antisocial traits [146]; high baseline depressive symptomatology [148]; type III and IV of Lesch's typology [148]; poor cognitive functioning [149]; Asn40Asp polymorphism of the μ -opioid receptor [150,151]; male gender [142,152]; type A of Babor's typology [153].

Of this, only Family History of Alcoholism and Asn40Asp polymorphism of the μ -opioid receptor has a consistent evidence of improving the effect of Naltrexone in patient with alcohol dependence [154]. However, the overall strength of evidence is weak, based on few studies with a high risk of bias.

NALMEFENE FOR THE TREATMENT OF ALCOHOL DEPENDENCE

Chemistry and Mechanism of Action

Nalmefene is a 6-methylene derivative of naltrexone, with high affinity at μ and κ -opioid receptors, and moderate affinity at δ -opioid receptor (Figure.6). It functions as an antagonist at μ and δ -opioid receptors and as partial agonist at κ -opioid receptors [155-158]. In one experimental study Nalmefene was found more effective than Naltrexone to suppressed alcohol self-administration in alcohol-dependent rats, but similar to Naltrexone in nondependent rats [159]. The differential binding profile of Nalmefene, and especially it's high affinity and partial agonist properties on κ -opioid receptors is advanced to explain this differential effect on ethanol dependent rats. Other beneficial differences relative to Naltrexone are: a better bioavailability [160], a longer plasma half-life [161], a longer occupancy of central μ -opioid receptors [161], resulting in a longer duration of action and no association with hepatotoxicity even with high doses [162].

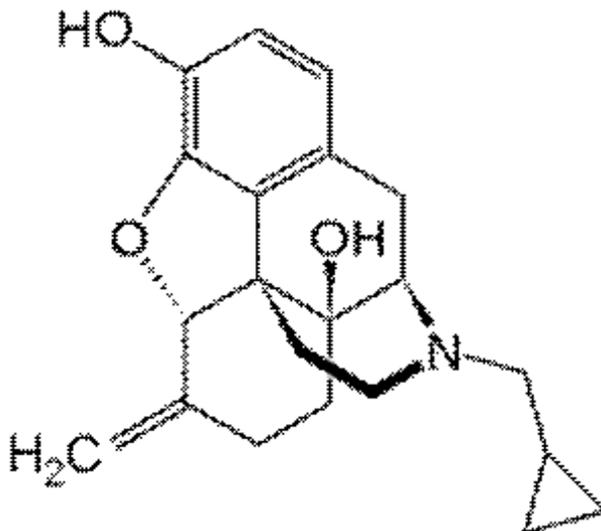


Figure 6: Chemical structure of Nalmefene.

Efficacy

Several studies evidenced the efficacy of Nalmefene for the treatment of alcoholism, but in a different fashion than Naltrexone, indeed in most studies the main goal was not abstinence but reduction of alcohol consumption with an as-needed administration of Nalmefene in patients who are not previously abstinent [163-166].

Three large multinational, randomized, double-blind, placebo-controlled, European trials support the efficacy of Nalmefene and led to its approval by the European Medicines Agency (EMA). ESSENSE 1 and ESSENSE 2 trials evaluated the efficacy of as-needed Nalmefene distributed to adults with alcohol dependence in reducing alcohol consumption in a 6-month period. ESSENSE 1 found that Nalmefene reduced the number of heavy drinking days (-2,3 day; $p=0,0021$) and total alcohol consumption (-11,0g/day; $p=0,0003$) [164]. ESSENSE 2 shows similar but less significant results, with a reduction of heavy drinking days (-1,7 day; $p=0,0,12$) and of total alcohol consumption (-5,0g/day; $p=0,088$) [165]. A post-hoc analysis of these two trials concluded that Nalmefene is more effective for patients with a high drinking risk level (men: >60 g/day; women: >40 g/day) [167].

SENSE trial evaluated the efficacy and safety of as-needed Nalmefene in a one-year period, concluding that Nalmefene is more effective than placebo in reducing the number of heavy drinking days (-1,6 day/month; $p=0,017$) and the total alcohol consumption (-6,5g/day; $p=0,036$) [166]. SENSE trial also concluded for the long-term safety of as-needed Nalmefene, evidencing mainly mild and transient adverse events. These 3 trials included a total of 1997 patients, and all included a motivational and adherence-enhancing intervention named BRENDA.

Management

Nalmefene (SELINCRO®) was approved by the EMA in 2013, for the reduction of alcohol consumption in alcohol dependent patients [168]. Contrary to Naltrexone, Nalmefene is to be taken on an as-needed basis. 1-2 hours preceding the perceived moment of drinking, the patient had to take one tablet of Nalmefene 18 mg, or as soon as possible if the patient had already start drinking. The drug is indicated for patients with alcohol dependence who have a high drinking risk level, without physical withdrawal symptoms and who do not require immediate detoxification. The marketing authorization also subline that Nalmefene should only be prescribed in conjunction with continuous psychosocial support focused on treatment adherence and reducing alcohol consumption. Finally, the drug should be initiated only in patients who continue to have a high drinking risk level two weeks after initial assessment. Nalmefene has already been integrated in French and NICE guidelines (133, 169), but is not already approved by the FDA for this indication.

Adverse Effects and Limitations of Use

Adverse effects are globally similar to that of Naltrexone including nausea, insomnia, dizziness, headache, fatigue and decreased appetite [166]. They are usually of mild and moderate intensity, and have a short time duration (1 to 7 days), associated with the treatment initiation. Contrary to Naltrexone high doses were not found to be associated with hepatotoxicity. A report of overdose with 450mg of Nalmefene didn't evidence alteration of vital functions [168]. As with Naltrexone, special cautions are to be taken with opioid addicts and patient taking medications containing an opioid.

Table 1: Comparison of Naltrexone and Nalmefene in alcohol dependence treatment.

	Naltrexone	Nalmefene
• Formula	$C_{20}H_{23}NO_4$	$C_{21}H_{25}NO_3$
• Trade name	REVIA®, DEPADE®, VIVITROL®	SELINCRO®
• Route of administration	Oral / Intra-muscular	Oral
• Oral bioavailability	~ 25%	~ 40%
• Half-life	~ 4h	~ 10h
• How to use in alcohol dependence	Initiate after detoxification, Orally: 50mg per day IM: 380mg every 4 weeks 6 month or more	Taken on an as-needed basis, 1-2h before the moment of drinking 18mg orally
• Aim	Maintaining abstinence	Reduction of alcohol consumption
• Adverse effects	nausea, vomiting, abdominal pain, fatigue, sleepiness risk of hepatotoxicity in high doses	nausea, insomnia, dizziness, headache, fatigue, decreased appetite
• Main vantages	Improved adhesion with depot formulation	Not associated with hepatotoxicity

Footnote

Although initially considered opioids antagonist, following studies demonstrated that Nalorphine and Cyclazocine are mixed opioid agonist/antagonist.

References

1. Gold MS. The neurobiology of addictive disorders: The role of dopamine, endorphin, and serotonin. The principles and practice of addictions in psychiatry Philadelphia: WB Saunders Company. 1997:57-69.
2. Di Chiara G, Imperato A, Mulas A. Preferential stimulation of dopamine release in the mesolimbic system: a common feature of drugs of abuse. Neurotransmitter interactions in the basal ganglia: Raven Press New York; 1987. 171-82.
3. Koob GF, Bloom FE. Cellular and molecular mechanisms of drug dependence. *Science*. 1988; 242: 715-723.
4. Lovinger DM, Grant KA. Alcohol neurotoxicity: Effects and mechanisms. *NEUROLOGICAL DISEASE AND THERAPY*. 1995; 36: 769.
5. Sellers EM, Higgins GA, Sobell MB. 5-HT and alcohol abuse. *Trends Pharmacol Sci*. 1992; 13: 69-75.
6. Woodward JJ. Alcohol. In: Miller NS, editor. Principles of addiction medicine: American Society of Addiction Medicine; 1994.
7. Fadda F, Mosca E, Colombo G, Gessa GL. Effect of spontaneous ingestion of ethanol on brain dopamine metabolism. *Life Sci*. 1989; 44: 281-287.
8. Gessa GL, Muntoni F, Collu M, Vargiu L, Mereu G. Low doses of ethanol activate dopaminergic neurons in the ventral tegmental area. *Brain Res*. 1985; 348: 201-203.
9. Yoshimoto K, Ueda S, Kato B, Takeuchi Y, Kawai Y. Alcohol enhances characteristic releases of dopamine and serotonin in the central nucleus of the amygdala. *Neurochem Int*. 2000; 37: 369-376.
10. Bustamante D, Quintanilla ME, Tampier L, Gonzalez-Lira V, Israel Y. Ethanol induces stronger dopamine release in nucleus accumbens (shell) of alcohol-preferring (bibulous) than in alcohol-avoiding (abstainer) rats. *Eur J Pharmacol*. 2008; 591: 153-158.
11. Blum K, Noble EP, Sheridan PJ, Montgomery A, Ritchie T. Allelic association of human dopamine D2 receptor gene in alcoholism. *JAMA*. 1990; 263: 2055-2060.
12. Pinto E, Reggers J, Gorwood P, Boni C, Scantamburlo G. The TaqI A DRD2 polymorphism in type II alcohol dependence: a marker of age at onset or of a familial disease? *Alcohol*. 2009; 43: 271-275.
13. Kraschewski A, Reese J, Anghelescu I, Winterer G, Schmidt LG, et al. Association of the dopamine D2 receptor gene with alcohol dependence: haplotypes and subgroups of alcoholics as key factors for understanding receptor function. *Pharmacogenetics and genomics*. 2009; 19: 513-527.
14. Kimura M, Higuchi S. Genetics of alcohol dependence. *Psychiatry Clin Neurosci*. 2011; 65: 213-225.
15. Setiawan E, Pihl RO, Dagher A, Schlagintweit H, Casey KF, et al. Differential striatal dopamine responses following oral alcohol in individuals at varying risk for dependence. *Alcoholism: Clinical and Experimental Research*. 2014; 38: 126-134.
16. Strother WN, Lumeng L, Li T-K, McBride WJ. Regional CNS densities of serotonin 1A and dopamine D 2 receptors in periadolescent alcohol-preferring P and alcohol-nonpreferring NP rat pups. *Pharmacology Biochemistry and Behavior*. 2003; 74: 335-342.
17. Volkow ND, Wang GJ, Fowler JS, Logan J, Hitzemann R. Decreases in dopamine receptors but not in dopamine transporters in alcoholics. *Alcohol Clin Exp Res*. 1996; 20: 1594-1598.
18. Tupala E, Hall H, Bergström K, Särkioja T, Räsänen P. Dopamine D(2)/D(3)-receptor and transporter densities in nucleus accumbens and amygdala of type 1 and 2 alcoholics. *Mol Psychiatry*. 2001; 6: 261-267.
19. Heinz A, Siessmeier T, Wrase J, Hermann D, Klein S, Grüsser-Sinopoli SM, et al. Correlation between dopamine D2 receptors in the ventral striatum and central processing of alcohol cues and craving. *American Journal of Psychiatry*. 2014.
20. Hietala J, West C, Syvälahti E, Nägren K, Lehtikoinen P. Striatal D2 dopamine receptor binding characteristics in vivo in patients with alcohol dependence. *Psychopharmacology (Berl)*. 1994; 116: 285-290.
21. Yoder KK, Kareken DA, Seyoum RA, O'connor SJ, Wang C. Dopamine D(2) receptor availability is associated with subjective responses to alcohol. *Alcohol Clin Exp Res*. 2005; 29: 965-970.
22. Volkow ND, Wang GJ, Begleiter H, Porjesz B, Fowler JS. High levels of dopamine D2 receptors in unaffected members of alcoholic families: possible protective factors. *Arch Gen Psychiatry*. 2006; 63: 999-1008.
23. Tupala E, Hall H, Mantere T, Räsänen P, Särkioja T, Tiihonen J. Dopamine receptors and transporters in the brain reward circuits of type 1 and 2 alcoholics measured with human whole hemisphere autoradiography?. *Neuroimage*. 2003; 19: 145-155.
24. Tampier L, Prado C, Quintanilla ME, Mardones J. Effect of bromocriptine on acute ethanol tolerance in UChB rats. *Addict Biol*. 1999; 4: 317-321.
25. Lawford BR, Young RM, Rowell JA, Qualichefski J, Fletcher BH. Bromocriptine in the treatment of alcoholics with the D2 dopamine receptor A1 allele. *Nat Med*. 1995; 1: 337-341.

26. Naranjo CA, Dongier M, Bremner KE. Long-acting injectable bromocriptine does not reduce relapse in alcoholics. *Addiction*. 1997; 92: 969-978.
27. Swift R. Medications acting on the dopaminergic system in the treatment of alcoholic patients. *Curr Pharm Des*. 2010; 16: 2136-2140.
28. Shaw GK, Waller S, Majumdar SK, Alberts JL, Latham CJ. Tiapride in the prevention of relapse in recently detoxified alcoholics. *Br J Psychiatry*. 1994; 165: 515-523.
29. Bender S, Scherbaum N, Soyka M, R  ther E, Mann K, et al. The efficacy of the dopamine D 2/D 3 antagonist tiapride in maintaining abstinence: a randomized, double-blind, placebo-controlled trial in 299 alcohol-dependent patients. *The International Journal of Neuropsychopharmacology*. 2007;10: 653-660.
30. Kishi T, Sevy S, Chekuri R, Correll CU. Antipsychotics for primary alcohol dependence: a systematic review and meta-analysis of placebo-controlled trials. *J Clin Psychiatry*. 2013; 74: e642-654.
31. Rossetti ZL, Melis F, Carboni S, Diana M, Gessa GL. Alcohol withdrawal in rats is associated with a marked fall in extraneuronal dopamine. *Alcoholism: Clinical and Experimental Research*. 1992;16(3):529-32.
32. Blum K, Cull JG, Braverman ER, Comings DE. Reward deficiency syndrome. *American Scientist*. 1996;84:132-145.
33. Trantham-Davidson H, Burnett EJ, Gass JT, Lopez MF, Mulholland PJ, et al. Chronic alcohol disrupts dopamine receptor activity and the cognitive function of the medial prefrontal cortex. *The Journal of Neuroscience*. 2014; 34: 3706-3718.
34. Sommer WH. Pathophysiology of alcohol addiction. In: Boyle P, Boffetta P, Lowenfels AB, Burns H, Brawley O, et al. editors. *Alcohol: Science, Policy and Public Health. Part II: OUP Oxford*. 2013; 84-97.
35. Pandley S, Davis, J Pandley G. Neurochemical findings in alcoholism and drug addiction and psychiatric comorbidity. In: Miller NS, editor. *The principles and practice of addictions in psychiatry: WB Saunders Company*; 1997; 70-79.
36. Gianoulakis C. Influence of the endogenous opioid system on high alcohol consumption and genetic predisposition to alcoholism. *Journal of psychiatry & neuroscience: JPN*. 2001; 26: 304.
37. Oswald LMW GS. Opioids and alcoholism. *Physiol Behav*. 2004; 81: 339-358.
38. Rada P, Barson JR, Leibowitz SF, Hoebel BG. Opioids in the hypothalamus control dopamine and acetylcholine levels in the nucleus accumbens. *Brain Res*. 2010; 1312: 1-9.
39. Froehlich JC. Opioid peptides. *Alcohol Health Res World*. 1997; 21: 132-136.
40. Nutt DJ. The role of the opioid system in alcohol dependence. *J Psychopharmacol*. 2014; 28: 8-22.
41. Lawrence Toll GC, Brian M Cox, Charles Chavkin, MacDonald J. Christie, Olivier Civelli, Mark Connor, Lakshmi A. Devi, Christopher Evans, Graeme Henderson, Volker H  llt, Brigitte Kieffer, Ian Kitchen, Mary-Jeanne Kreek, Lee-Yuan Liu-Chen, Jean-Claude Meunier, Philip S. Portoghese, Toni S. Shippenberg, Eric J Simon, John R Traynor, Hiroshi Ueda, Yung H. Wong. Opioid receptors, introduction: IUPHAR/BPS Guide to PHARMACOLOGY; 2015.
42. McNally GP, Akil H. *3opioid Peptides and Their Receptors: Overview and Function in Pain Modulation*. 2002.
43. Noble F, Benturquia N, Bilkei-Gorzo A, Zimmer A, Roques BP. Use of preproenkephalin knockout mice and selective inhibitors of enkephalinases to investigate the role of enkephalins in various behaviours. *Psychopharmacology (Berl)*. 2008; 196: 327-335.
44. Fichna J, Janecka A, Costentin J, Do Rego JC. The endomorphin system and its evolving neurophysiological role. *Pharmacol Rev*. 2007; 59: 88-123.
45. Koneru A, Satyanarayana S, Rizwan S. Endogenous opioids: their physiological role and receptors. *Global J Pharmacol*. 2009; 3:149-153.
46. Barry SM, Grisel JE. * -Endorphin and Alcoholism: INTECH Open Access Publisher*; 2012.
47. Veening JG, Gerrits PO, Barendregt HP. Volume transmission of beta-endorphin via the cerebrospinal fluid; a review. *Fluids Barriers CNS*. 2012; 9: 16.
48. Schwarzer C. 30 years of dynorphins--new insights on their functions in neuropsychiatric diseases. *Pharmacol Ther*. 2009; 123: 353-370.
49. Asvadi NH, Morgan M, Hewavitharana AK, Shaw PN, Cabot PJ. Biotransformation of beta-endorphin and possible therapeutic implications. *Front Pharmacol*. 2014; 5: 18.
50. Momeni S. Individual differences in behavior, neurochemistry and pharmacology associated with voluntary alcohol intake. 2015.
51. Martins RT, de Almeida DB, do Rego Monteiro FM, Andr   P, Kowacs RR. Receptores opioides at   o contexto atual. *Revista Dor*. 2012; 13: 75-84.

52. Kieffer BL, Evans CJ. Opioid receptors: from binding sites to visible molecules in vivo. *Neuropharmacology*. 2009; 56: 205-212.
53. Devine DP, Leone P, Pocock D, Wise RA. Differential involvement of ventral tegmental mu, delta and kappa opioid receptors in modulation of basal mesolimbic dopamine release: in vivo microdialysis studies. *J Pharmacol Exp Ther*. 1993; 266: 1236-1246.
54. Kranzler HR, Anton RF. Implications of recent neuropsychopharmacologic research for understanding the etiology and development of alcoholism. *Journal of consulting and clinical psychology*. 1994;62(6):1116.
55. Grisel JE, Bartels JL, Allen SA, Turgeon VL. Influence of beta-Endorphin on anxious behavior in mice: interaction with EtOH. *Psychopharmacology (Berl)*. 2008; 200: 105-115.
56. De Waele JP, Papachristou DN, Gianoulakis C. The alcohol-preferring C57BL/6 mice present an enhanced sensitivity of the hypothalamic beta-endorphin system to ethanol than the alcohol-avoiding DBA/2 mice. *J Pharmacol Exp Ther*. 1992; 261: 788-794.
57. Marinelli PW, Lam M, Bai L, Quirion R, Gianoulakis C. A microdialysis profile of dynorphin A1–8 release in the rat nucleus accumbens following alcohol administration. *Alcoholism: Clinical and Experimental Research*. 2006; 30: 982-990.
58. Rasmussen DD, Boldt BM, Wilkinson CW, Mitton DR. Chronic Daily Ethanol and Withdrawal: 3. Forebrain Pro-Opiomelanocortin Gene Expression and Implications for Dependence, Relapse, and Deprivation Effect. *Alcoholism: Clinical and Experimental Research*. 2002; 26: 535-546.
59. D'Addario C, Caputi FF, Rimondini R, Gandolfi O, Del Borrello E, et al. Different alcohol exposures induce selective alterations on the expression of dynorphin and nociceptin systems related genes in rat brain. *Addiction biology*. 2013; 18: 425-433.
60. Nylander I, Hyttiä P, Forsander O, Terenius L. Differences between Alcohol-Preferring (AA) and Alcohol-Avoiding (ANA) Rats in the Prodynorphin and Proenkephalin Systems. *Alcoholism: Clinical and Experimental Research*. 1994;18: 1272-1279.
61. Chang GQ, Barson JR, Karatayev O, Chang SY, Chen YW. Effect of Chronic Ethanol on Enkephalin in the Hypothalamus and Extra-Hypothalamic Areas. *Alcoholism: Clinical and Experimental Research*. 2010; 34: 761-770.
62. De Waele JP, Gianoulakis C. Enhanced activity of the brain beta-endorphin system by free-choice ethanol drinking in C57BL/6 but not DBA/2 mice. *Eur J Pharmacol*. 1994; 258: 119-129.
63. Marinelli PW, Kiianmaa K, Gianoulakis C. Opioid propeptide mRNA content and receptor density in the brains of AA and ANA rats. *Life Sci*. 2000; 66: 1915-1927.
64. Gianoulakis C, Waele JP, Kiianmaa K. Differences in the Brain and Pituitary β -Endorphin System between the Alcohol-Preferring AA and Alcohol-Avoiding ANA Rats. *Alcoholism: Clinical and Experimental Research*. 1992; 16: 453-459.
65. Krishnan-Sarin S, Wand GS, Li XW, Portoghese PS, Froehlich JC. Effect of mu opioid receptor blockade on alcohol intake in rats bred for high alcohol drinking. *Pharmacology Biochemistry and Behavior*. 1998; 59: 627-635.
66. Karatayev O, Barson JR, Carr AJ, Baylan J, Chen Y-W, Leibowitz SF. Predictors of ethanol consumption in adult Sprague–Dawley rats: relation to hypothalamic peptides that stimulate ethanol intake. *Alcohol*. 2010; 44: 323-334.
67. Ng GY, O Dowd BF, George SR. Genotypic differences in mesolimbic enkephalin gene expression in DBA/2J and C57BL/6J inbred mice. *Eur J Pharmacol*. 1996; 311: 45-52.
68. Li XW, Li TK, Froehlich JC. Enhanced sensitivity of the nucleus accumbens proenkephalin system to alcohol in rats selectively bred for alcohol preference. *Brain Res*. 1998; 794: 35-47.
69. Jamensky NT, Gianoulakis C. Content of dynorphins and kappa-opioid receptors in distinct brain regions of C57BL/6 and DBA/2 mice. *Alcohol Clin Exp Res*. 1997; 21: 1455-1464.
70. McBride WJ, Chernet E, McKinzie DL, Lumeng L, Li TK. Quantitative autoradiography of mu-opioid receptors in the CNS of alcohol-naive alcohol-preferring P and -nonpreferring NP rats. *Alcohol*. 1998; 16: 317-323.
71. Waele JP, Gianoulakis C. Characterization of the μ and δ opioid receptors in the brain of the C57BL/6 and DBA/2 mice, selected for their differences in voluntary ethanol consumption. *Alcoholism: Clinical and Experimental Research*. 1997; 21: 754-762.
72. Roberts AJ, McDonald JS, Heyser CJ, Kieffer BL, Matthes HW. mu-Opioid receptor knockout mice do not self-administer alcohol. *J Pharmacol Exp Ther*. 2000; 293: 1002-1008.
73. Gianoulakis C, Béliveau D, Angelogianni P, Meaney M, Thavundayil J. Different pituitary beta-endorphin and adrenal cortisol response to ethanol in individuals with high and low risk for future development of alcoholism. *Life Sci*. 1989; 45: 1097-1109.
74. Gianoulakis C, Krishnan B, Thavundayil J. Enhanced sensitivity of pituitary beta-endorphin to ethanol in subjects at high risk of alcoholism. *Arch Gen Psychiatry*. 1996; 53: 250-257.
75. Dai X, Thavundayil J, Gianoulakis C. Differences in the peripheral levels of beta-endorphin in response to alcohol and stress as a function of alcohol dependence and family history of alcoholism. *Alcohol Clin Exp Res*. 2005; 29: 1965-1975.

76. Genazzani AR, Nappi G, Facchinetti F, Mazzella GL, Parrini D. Central deficiency of beta-endorphin in alcohol addicts. *J Clin Endocrinol Metab.* 1982; 55: 583-586.
77. Aguirre JC, Del Arbol JL, Raya J, Ruiz-Requena ME, Rico Irlas J. Plasma beta-endorphin levels in chronic alcoholics. *Alcohol.* 1990; 7: 409-412.
78. Mitchell JM, O'Neil JP, Janabi M, Marks SM, Jagust WJ. Alcohol consumption induces endogenous opioid release in the human orbitofrontal cortex and nucleus accumbens. *Science Translational Medicine.* 2012; 4: ra6.
79. Ulm RR, Volpicelli JR, Volpicelli LA. Opiates and alcohol self-administration in animals. *J Clin Psychiatry.* 1995; 56: 5-14.
80. Trachtenberg MC, Blum K. Alcohol and opioid peptides: neuropharmacological rationale for physical craving of alcohol. *Am J Drug Alcohol Abuse.* 1987; 13: 365-372.
81. Reid LD, Delconte JD, Nichols ML, Bilsky EJ, Hubbell CL. Tests of opioid deficiency hypotheses of alcoholism. *Alcohol.* 1991; 8: 247-257.
82. McCubbin JA. Stress and endogenous opioids: behavioral and circulatory interactions. *Biol Psychol.* 1993; 35: 91-122.
83. Lee S, Selvage D, Hansen K, Rivier C. Site of action of acute alcohol administration in stimulating the rat hypothalamic-pituitary-adrenal axis: comparison between the effect of systemic and intracerebroventricular injection of this drug on pituitary and hypothalamic responses. *Endocrinology.* 2004; 145: 4470-4479.
84. Costa A, Bono G, Martignoni E, Merlo P, Sances G. An assessment of hypothalamo-pituitary-adrenal axis functioning in non-depressed, early abstinent alcoholics. *Psychoneuroendocrinology.* 1996; 21: 263-275.
85. Zimmermann U, Spring K, Kunz-Ebrecht SR, Uhr M, Wittchen HU. Effect of ethanol on hypothalamic-pituitary-adrenal system response to psychosocial stress in sons of alcohol-dependent fathers. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology.* 2004; 29: 1156-1165.
86. Zalewska-Kaszubska J, Czamecka E. Deficit in beta-endorphin peptide and tendency to alcohol abuse. *Peptides.* 2005; 26: 701-705.
87. Adinoff B, Junghanns K, Kiefer F, Krishnan-Sarin S. Suppression of the HPA axis stress-response: implications for relapse. *Alcoholism, clinical and experimental research.* 2005; 29: 1351-1355.
88. Kiefer F, Horntrich M, Jahn H, Wiedemann K. Is withdrawal-induced anxiety in alcoholism based on beta-endorphin deficiency? *Psychopharmacology (Berl).* 2002; 162: 433-437.
89. Goldowitz D, Matthews DB, Hamre KM, Mittleman G, Chesler EJ, et al. Progress in using mouse inbred strains, consomics, and mutants to identify genes related to stress, anxiety, and alcohol phenotypes. *Alcoholism, clinical and experimental research.* 2006; 30: 1066-1078.
90. Hubbell CL, Czirr SA, Hunter GA, Beaman CM, LeCann NC. Consumption of ethanol solution is potentiated by morphine and attenuated by naloxone persistently across repeated daily administrations. *Alcohol.* 1986; 3: 39-54.
91. Benjamin D, Grant ER, Pohorecky LA. Naltrexone reverses ethanol-induced dopamine release in the nucleus accumbens in awake, freely moving rats. *Brain Res.* 1993; 621: 137-140.
92. Heyser CJ, Roberts AJ, Schulteis G, Koob GF. Central administration of an opiate antagonist decreases oral ethanol self-administration in rats. *Alcohol Clin Exp Res.* 1999; 23: 1468-1476.
93. O'Malley SS, Jaffe AJ, Chang G, Schottenfeld RS, Meyer RE. Naltrexone and coping skills therapy for alcohol dependence. A controlled study. *Arch Gen Psychiatry.* 1992; 49: 881-887.
94. Volpicelli JR, Alterman AI, Hayashida M, O'Brien CP. Naltrexone in the treatment of alcohol dependence. *Arch Gen Psychiatry.* 1992; 49: 876-880.
95. Sinclair JD. Evidence about the use of naltrexone and for different ways of using it in the treatment of alcoholism. *Alcohol Alcohol.* 2001; 36: 2-10.
96. Chick J, Anton R, Checinski K, Croop R, Drummond DC. A multicentre, randomized, double-blind, placebo-controlled trial of naltrexone in the treatment of alcohol dependence or abuse. *Alcohol Alcohol.* 2000; 35: 587-593.
97. Mason BJ, Salvato FR, Williams LD, Ritvo EC, Cutler RB. A double-blind, placebo-controlled study of oral nalmefene for alcohol dependence. *Arch Gen Psychiatry.* 1999; 56: 719-724.
98. Pohl J. Über das N-allylnorcodein, einen Antagonisten des Morphins. *Zeitschrift für experimentelle Pathologie und Therapie.* 1915; 17: 370-382.
99. Hart E. N-allylnorcodeine and N-allylnormorphine, two antagonists to morphine. *J Pharmacol Exp Ther.* 1941; 72: 356-363.
100. HART ER, McCRAWLEY EL. The pharmacology of N-allylnormorphine as compared with morphine. *Journal of Pharmacology and Experimental Therapeutics.* 1944; 82: 339-348.

101. Eckenhoff J, Elder J, King B. The effect of N-allyl normorphine in treatment of opiate overdose. AMERICAN JOURNAL OF THE MEDICAL SCIENCES; 1951: LIPPINCOTT WILLIAMS & WILKINS 227 EAST WASHINGTON SQ, PHILADELPHIA, PA 19106.
102. Lasagna L, Beecher Hk. The analgesic effectiveness of codeine and meperidine (demerol). J Pharmacol Exp Ther. 1954; 112: 306-311.
103. Archer S, Albertson NF, Harris LS, Pierson AK, Bird JG. Narcotic antagonists as analgesics. Science. 1962; 137: 541-543.
104. Harris LS, Pierson AK. Some Narcotic Antagonists in The Benzomorphan Series. J Pharmacol Exp Ther. 1964; 143: 141-148.
105. Martin W, Gorodetzky C, McClane T. A proposed method for ambulatory treatment of narcotic addicts using a long-acting, orally effective narcotic antagonist cyclazocine: An experimental study. Committee on Problems of Drug Dependence. 1965.
106. Martin WR, Gorodetzky CW, McClane TK. An experimental study in the treatment of narcotic addicts with cyclazocine. Clin Pharmacol Ther. 1966; 7: 455-465.
107. Freedman AM, Fink M, Sharoff R, Zaks A. Clinical studies of cyclazocine in the treatment of narcotic addiction. Am J Psychiatry. 1968; 124: 1499-1504.
108. Bidlack JM, Cohen DJ, McLaughlin JP, Lou R, Ye Y. 8-Carboxamidocyclazocine: a long-acting, novel benzomorphan. J Pharmacol Exp Ther. 2002; 302: 374-380.
109. Van Dorp E, Yassen A, Dahan A. Naloxone treatment in opioid addiction: the risks and benefits. Expert Opin Drug Saf. 2007; 6: 125-132.
110. Jasinski DR, Martin WR, Haertzen CA. The human pharmacology and abuse potential of N-allylnoroxymorphone (naloxone). J Pharmacol Exp Ther. 1967; 157: 420-426.
111. Fink M, Zaks A, Sharoff R, Mora A, Bruner A. Naloxone in heroin dependence. Clin Pharmacol Ther. 1968; 9: 568-577.
112. Ngai SH, Berkowitz BA, Yang JC, Hempstead J, Spector S. Pharmacokinetics of naloxone in rats and in man: basis for its potency and short duration of action. Anesthesiology. 1976; 44: 398-401.
113. Blumberg H, Pachter I, Matossian Z. Patent 3 332 950, 1967 Chem. Abstr; 1967.
114. Blumberg H, Dayton H, Wolf P. Analgesic and narcotic antagonist properties of noroxymorphone derivatives. Toxicology and Applied Pharmacology; 1967: ACADEMIC PRESS INC 525 B ST, STE 1900, SAN DIEGO, CA 92101-4495.
115. Martin W, Jasinski D. Characterization of n-cyclopropylmethyl-7, 8-dihydro-14-hydroxynormorphine hcl, a narcotic antagonist. Clinical pharmacology & therapeutics; 1973: mosby-year book inc 11830 westline industrial dr, st louis, mo 63146-63318.
116. Codd EE, Shank RP, Schupsky JJ, Raffa RB. Serotonin and norepinephrine uptake inhibiting activity of centrally acting analgesics: structural determinants and role in antinociception. J Pharmacol Exp Ther. 1995; 274: 1263-1270.
117. Marfaing-Jallat P, Miceli D, Le Magnen J. Decrease in ethanol consumption by naloxone in naive and dependent rats. Pharmacol Biochem Behav. 1983; 18 Suppl 1: 537-539.
118. Samson HH, Doyle TF. Oral ethanol self-administration in the rat: effect of naloxone. Pharmacol Biochem Behav. 1985; 22: 91-99.
119. Froehlich J, Harts J, Lumeng L, Li T-K. Naloxone attenuates voluntary ethanol intake in rats selectively bred for high ethanol preference. Pharmacology Biochemistry and Behavior. 1990;35: 385-390.
120. Martin WR, Jasinski DR, Mansky PA. Naltrexone, an antagonist for the treatment of heroin dependence. Effects in man. Arch Gen Psychiatry. 1973; 28: 784-791.
121. Verebey K, Mule S. Naltrexone pharmacology, pharmacokinetics, and metabolism: current status. The American journal of drug and alcohol abuse. 1975; 2: 357-363.
122. Wentland MP, Lou R, Lu Q, Bu Y, Denhardt C. Syntheses of novel high affinity ligands for opioid receptors. Bioorg Med Chem Lett. 2009; 19: 2289-2294.
123. Weerts EM, Kim YK, Wand GS, Dannals RF, Lee JS, Frost JJ, et al. Differences in d- and μ -opioid receptor blockade measured by positron emission tomography in naltrexone-treated recently abstinent alcohol-dependent subjects. Neuropsychopharmacology. 2008; 33: 653-665.
124. Johnson BA. Update on neuropharmacological treatments for alcoholism: scientific basis and clinical findings. Biochemical pharmacology. 2008; 75: 34-56.
125. Verebey K, Volavka J, Mulé SJ, Resnick RB. Naltrexone: disposition, metabolism, and effects after acute and chronic dosing. Clin Pharmacol Ther. 1976; 20: 315-328.
126. McCaul ME, Wand GS, Rohde C, Lee SM. Serum 6-beta-naltrexol levels are related to alcohol responses in heavy drinkers. Alcohol Clin Exp Res. 2000; 24: 1385-1391.

127. Anton RF, O'Malley SS, Ciraulo DA, Cisler RA, Couper D. Combined pharmacotherapies and behavioral interventions for alcohol dependence: the COMBINE study: a randomized controlled trial. *JAMA*. 2006; 295: 2003-2017.
128. Donovan DM, Anton RF, Miller WR, Longabaugh R, Hosking JD. Combined pharmacotherapies and behavioral interventions for alcohol dependence (The COMBINE Study): examination of posttreatment drinking outcomes. *J Stud Alcohol Drugs*. 2008; 69: 5-13.
129. Rösner S, Hackl-Herrwerth A, Leucht S, Vecchi S, Srisurapanont M, Soyka M. Opioid antagonists for alcohol dependence. *Cochrane Database Syst Rev*. 2010; 12.
130. Treatment CfSA. Incorporating Alcohol Pharmacotherapies Into Medical Practice: A Review of the Literature. 2009.
131. Health NCCfM, Health Nif, Excellence C. Alcohol-use disorders: diagnosis, assessment and management of harmful drinking and alcohol dependence: RCPsych Publications; 2011.
132. Strain EC. Incorporating Alcohol Pharmacotherapies Into Medical Practice: A Treatment Improvement Protocol: DIANE Publishing; 2010.
133. Aubin PH-J, Gillet C, Rigaud A. Mésusage de l'alcool dépitage, diagnostic et traitement. *Alcoologie et Addictologie*. 2015; 37: 5-84.
134. Mitchell JE, Morley JE, Levine AS, Hatsukami D, Gannon M, Pfohl D. High-dose naltrexone therapy and dietary counseling for obesity. *Biological psychiatry*. 1987; 22: 35-42.
135. Atkinson RL, Berke LK, Drake CR, Bibbs ML, Williams FL, Kaiser DL. Effects of long-term therapy with naltrexone on body weight in obesity. *Clinical Pharmacology & Therapeutics*. 1985; 38: 419-422.
136. Chick J, Anton R, Checinski K, Croop R, Drummond DC, Farmer R, et al. A multicentre, randomized, double-blind, placebo-controlled trial of naltrexone in the treatment of alcohol dependence or abuse. *Alcohol and alcoholism*. 2000; 35: 587-593.
137. Volpicelli JR, Rhines KC, Rhines JS, Volpicelli LA, Alterman AI, O'Brien CP. Naltrexone and alcohol dependence: role of subject compliance. *Archives of General Psychiatry*. 1997; 54: 737-742.
138. Gueorguieva R, Wu R, Pittman B, Cramer J, Rosenheck RA, O'Malley SS, et al. New insights into the efficacy of naltrexone based on trajectory-based reanalyses of two negative clinical trials. *Biological psychiatry*. 2007; 61: 1290-1295.
139. Kranzler HR, Stephenson JJ, Montejano L, Wang S, Gastfriend DR. Persistence with oral naltrexone for alcohol treatment: implications for health-care utilization. *Addiction*. 2008; 103: 1801-1808.
140. Harris KM, DeVries A, Dimidjian K, Pincus H, Tanielian T. Trends in naltrexone use among members of a large private health plan. *Psychiatr Serv*. 2004; 55: 221.
141. Dunbar JL, Turncliff RZ, Dong Q, Silverman BL, Ehrich EW, Lasseter KC. Single-and Multiple-Dose Pharmacokinetics of Long-acting Injectable Naltrexone. *Alcoholism: Clinical and Experimental Research*. 2006; 30: 480-490.
142. Garbutt JC, Kranzler HR, O'Malley SS, Gastfriend DR, Pettinati HM, Silverman BL, et al. Efficacy and tolerability of long-acting injectable naltrexone for alcohol dependence: a randomized controlled trial. *Jama*. 2005; 293: 1617-1625.
143. O'Malley SS, Garbutt JC, Gastfriend DR, Dong Q, Kranzler HR. Efficacy of extended-release naltrexone in alcohol-dependent patients who are abstinent before treatment. *Journal of clinical psychopharmacology*. 2007; 27: 507-512.
144. Rubio G, Ponce G, Rodriguez-Jimenez R, Jimenez-Arriero M, Hoenicka J, Palomo T. Clinical predictors of response to naltrexone in alcoholic patients: who benefits most from treatment with naltrexone? *Alcohol and Alcoholism*. 2005; 40: 227-233.
145. Monterosso JR, Flannery BA, Pettinati HM, Oslin DW, Rukstalis M, O'Brien CP, et al. Predicting treatment response to naltrexone: the influence of craving and family history. *The American Journal on Addictions*. 2001; 10: 258-268.
146. Rohsenow DJ, Miranda Jr R, McGeary JE, Monti PM. Family history and antisocial traits moderate naltrexone's effects on heavy drinking in alcoholics. *Experimental and clinical psychopharmacology*. 2007; 15: 272.
147. Krishnan-Sarin S, Krystal JH, Shi J, Pittman B, O'Malley SS. Family history of alcoholism influences naltrexone-induced reduction in alcohol drinking. *Biological psychiatry*. 2007; 62: 694-697.
148. Kiefer F, Helwig H, Tarnaske T, Otte C, Jahn H, Wiedemann K. Pharmacological relapse prevention of alcoholism: clinical predictors of outcome. *European addiction research*. 2005; 11: 83-91.
149. Jaffe AJ, Rounsaville B, Chang G, Schottenfeld RS, Meyer RE, O'Malley SS. Naltrexone, relapse prevention, and supportive therapy with alcoholics: an analysis of patient treatment matching. *Journal of consulting and clinical psychology*. 1996; 64: 1044.
150. Jonas DE, Amick HR, Feltner C, Wines R, Shanahan E, Rowe CJ, et al. Genetic polymorphisms and response to medications for alcohol use disorders: a systematic review and meta-analysis. *Pharmacogenomics*. 2014; 15: 1687-700.

151. Chamorro AJ, Marcos M, Mirón-Canelo JA, Pastor I, González-Sarmiento R, Laso FJ. Association of μ -opioid receptor (OPRM1) gene polymorphism with response to naltrexone in alcohol dependence: a systematic review and meta-analysis. *Addiction biology*. 2012; 17: 505-512.
152. Kranzler HR, Tennen H, Armeli S, Chan G, Covault J, Arias A, et al. Targeted naltrexone for problem drinkers. *Journal of clinical psychopharmacology*. 2009; 29: 350.
153. Bogenschütz MP, Scott Tonigan J, Pettinati HM. Effects of alcoholism typology on response to naltrexone in the COMBINE study. *Alcoholism: Clinical and Experimental Research*. 2009; 33: 10-18.
154. Garbutt JC, Greenblatt AM, West SL, Morgan LC, Kampov-Polevoy A, Jordan HS, et al. Clinical and biological moderators of response to naltrexone in alcohol dependence: a systematic review of the evidence. *Addiction*. 2014; 109: 1274-1284.
155. Michel M, Bolger G, Weissman B. Binding of a new opiate antagonist, nalmefene, to rat brain membranes. *Methods and findings in experimental and clinical pharmacology*. 1985; 7: 175-177.
156. Bart G, Schluger JH, Borg L, Ho A, Bidlack JM, Kreek MJ. Nalmefene induced elevation in serum prolactin in normal human volunteers: partial kappa opioid agonist activity? *Neuropsychopharmacology*. 2005; 30: 2254-2262.
157. Osborn MD, Lowery JJ, Skorput AG, Giuvelis D, Bilsky EJ. In vivo characterization of the opioid antagonist nalmefene in mice. *Life sciences*. 2010; 86: 624-630.
158. Nealey KA, Smith AW, Davis SM, Smith DG, Walker BM. κ -opioid receptors are implicated in the increased potency of intracumbens nalmefene in ethanol-dependent rats. *Neuropharmacology*. 2011; 61: 35-42.
159. Walker BM, Koob GF. Pharmacological evidence for a motivational role of κ -opioid systems in ethanol dependence. *Neuropsychopharmacology*. 2008; 33: 643-652.
160. Broksø Kuhl LE, Li S, Faerch KU, Soegaard B, Larsen F. Population Pharmacokinetics of Nalmefene in Healthy Subjects and its Relation to μ -Opioid Receptor Occupancy. *British journal of clinical pharmacology*. 2015.
161. Ingman K, Hagelberg N, Aalto S, Någren K, Juhakoski A, et al. Prolonged central μ -opioid receptor occupancy after single and repeated nalmefene dosing. *Neuropsychopharmacology*. 2005; 30: 2245-2253.
162. Mason BJ, Salvato FR, Williams LD, Ritvo EC, Cutler RB. A double-blind, placebo-controlled study of oral nalmefene for alcohol dependence. *Archives of General Psychiatry*. 1999; 56: 719-724.
163. Karhuvaara S, Simojoki K, Virta A, Rosberg M, Löyttyniemi E, et al. Targeted nalmefene with simple medical management in the treatment of heavy drinkers: a randomized double-blind placebo-controlled multicenter study. *Alcoholism: Clinical and Experimental Research*. 2007; 31: 1179-1187.
164. Mann K, Bladström A, Torup L, Gual A, Van den Brink W. Extending the treatment options in alcohol dependence: a randomized controlled study of as-needed nalmefene. *Biological psychiatry*. 2013; 73: 706-713.
165. Gual A, He Y, Torup L, Van den Brink W, Mann K. A randomised, double-blind, placebo-controlled, efficacy study of nalmefene, as-needed use, in patients with alcohol dependence. *European neuropsychopharmacology*. 2013; 23: 1432-1442.
166. Van den Brink W, Sørensen P, Torup L, Mann K, Gual A, Group SS. Long-term efficacy, tolerability and safety of nalmefene as-needed in patients with alcohol dependence: A 1-year, randomised controlled study. *Journal of Psychopharmacology*. 2014:0269881114527362.
167. Van den Brink W, Aubin H-J, Bladström A, Torup L, Gual A, Mann K. Efficacy of as-needed nalmefene in alcohol-dependent patients with at least a high drinking risk level: results from a subgroup analysis of two randomized controlled 6-month studies. *Alcohol and alcoholism*. 2013; 48: 570-578.
168. Lundbeck A. Nalmefene (Selincro) [summary of product characteristics]. www.ema.europa.eu. 2013. 2014.
169. Stevenson M, Pandor A, Stevens JW, Rawdin A, Rice P, et al. Nalmefene for reducing alcohol consumption in people with alcohol dependence: an evidence review group perspective of a NICE single technology appraisal. *Pharmacoeconomics*. 2015; 33: 833-847.