

Duality and Plasticity of Th17 Cells in Behçet's Disease

Kamel Hamzaoui^{1*} and Agnes Hamzaoui²

¹Department of Basic Sciences, University of Tunis El Manar, Tunisia

²Department of Paediatric Respiratory Diseases, University of Tunis El Manar, Tunisia

***Corresponding author:** Kamel Hamzaoui, University of Tunis El Manar, Tunisia, Tel: +216 98 325 181; Email: kamel.hamzaoui@gmail.com

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ABSTRACT

T helper type 17 (Th17) cells are active players in the establishment of inflammation in Behçet's disease (BD). How this process impacts the immune dysregulation observed in BD is far from fully understood. Plasticity within this subset is suggested by the existence of IL-17 secreting cells, which express either forkhead box p3 (Foxp3) or retinoic acid receptor-related orphan nuclear receptor (ROR γ t), signature transcription factors of Treg and Th17 respectively. Much more is unknown at the basic level, as IL-17 producing cells do not always contribute to inflammation, and may even acquire regulatory features depending upon environment modifications. This review aims to highlight Th17 plasticity and to discuss the role of Th17/IL-17-axis in BD inflammatory pathway.

Keywords: Behçet's disease; Th17; Inflammation; ROR γ t; Duality; Plasticity; BTLA

INTRODUCTION

CD4+ T cells' ability to exert their effector functions depends on the provision from antigen-presenting cells (dendritic cells: DC) of the immunological cues that prompt formation of Th subsets: Th1, Th2, Th9, Th17, Th22, and FoxP3+ regulatory T (Treg) cells (Figure 1) [1-3]. Th17 subset is so named due to its ability to secrete interleukin (IL)-17A, which has emerged as a major player in tissue-specific immune pathology, interacting with Th1 and Treg lymphocytes. The initial emphasis on the detrimental cytotoxic effects of Th17 is reflected by the plethora of early literature supporting such a role in both human and murine studies [2-4].

Th17 cells and their associated cytokines have been found to interact closely with other adaptive immune cells, raising interesting questions about how to select and design therapeutic strategies targeting this cell population [5]. New technologies such as transcriptome profiling, global epigenetic mapping and computerized simulation analysis [6-7] have captured an accurate picture of this T cell subset, revealing it to be more transient, complex and perhaps more reversible than previously imagined. Human Foxp3+ Treg cells can differentiate into IL-17 promoting cells *in vitro* [8]. Flexibility within this subset may allow Th17 cells to embrace either pro-inflammatory or protective roles in inflammatory and auto-inflammatory diseases by secreting a wide spectrum of cytokines (Figure 1).

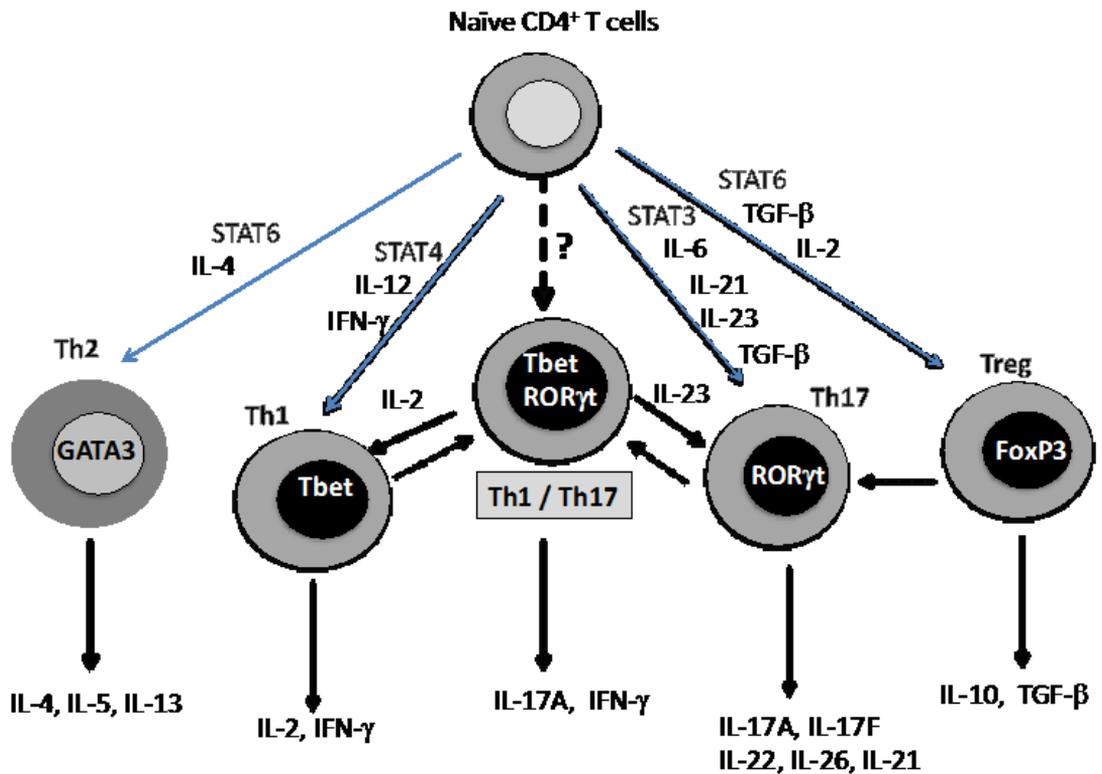


Figure 1: Cytokines and transcription factors required for helper (Th) differentiation.

In the presence of interleukin-6 (IL-6), IL-21, and transforming growth factor-beta (TGF-β), naïve CD4⁺ T cells differentiate into a Th17 cell phenotype, which is characterized by the expression of transcription factors retinoic acid receptor-related orphan receptor-γt (RORγt) and signal transducer and activator of transcription 3 (STAT3). IL-1β and IL-23 cytokines can promote and stabilize this phenotype during cell expansion. Once programmed, these cells secrete IL-17A, IL-17F, IL-21, and IL-22, which play a key role in enhancing autoimmunity and host defence. The differentiation of Th17 cells is promoted by activation of STAT3 and inhibited by activation of STAT1. Cytokines IL-12, IL-4, and TGF-β and transcription factors T-bet, GATA3, and FoxP3 have been shown to regulate Th1, Th2, and Treg cell development, respectively.

These distinct subsets regulate immune response. (R): cytokine receptor.

In BD, an imbalance between Th17 cells and Treg cells may be generated quickly in order to establish immune homeostasis [9-11]. This ability to transition between functional states is defined as T cell plasticity [12]. Differentiated CD4⁺ T cell subpopulations display a high grade of plasticity. Their initial differentiation is not an endpoint of T cell development that allows a functional adaptation to various physiological situations during immune response. Plasticity of effector helper T cells offers the opportunity, for clinical immunologists, to interfere with the natural course of immune-mediated diseases. This could be possible either by blocking

environmental signals that drive the transition of T helper cells towards more aggressive phenotypes, or by promoting the differentiation towards less pathogenic phenotypes [13].

This review outlines the major features of Th17 plasticity including the Treg/Th17 paradigm and discusses this duality in Behçet's disease focusing on the maintenance of homeostasis.

TH17 LINEAGE: IL-17 AND IL-17 PRODUCING CELLS

Th17 cells are characterized by secretion of IL-17A, expression of chemokine receptor CCR6 and transcriptional factor ROR γ t [14]. Th17 pathogenicity is limited by Foxp3⁺ Treg and T regulatory type 1 (TR1) cells [14-15]. Treg cells are characterized by the transcription factor Foxp3, whereas TR1 cells secrete high levels of the anti-inflammatory IL-10 and express cell-surface markers CD49b [15-17]. Although Th17, Foxp3⁺ Treg and TR1 cells are functionally distinct subsets, they share several features. Their differentiation is promoted by transforming growth factor β (TGF- β) [18] and both Th17 and TR1 cells express CD49b and high levels of the transcription factor aryl hydrocarbon receptor 9 (AhR9). Moreover Th17 cells can transiently co-express ROR γ t with Foxp3 [19-20] and IL-17A with IL-10 [21-23] (Figure 2).

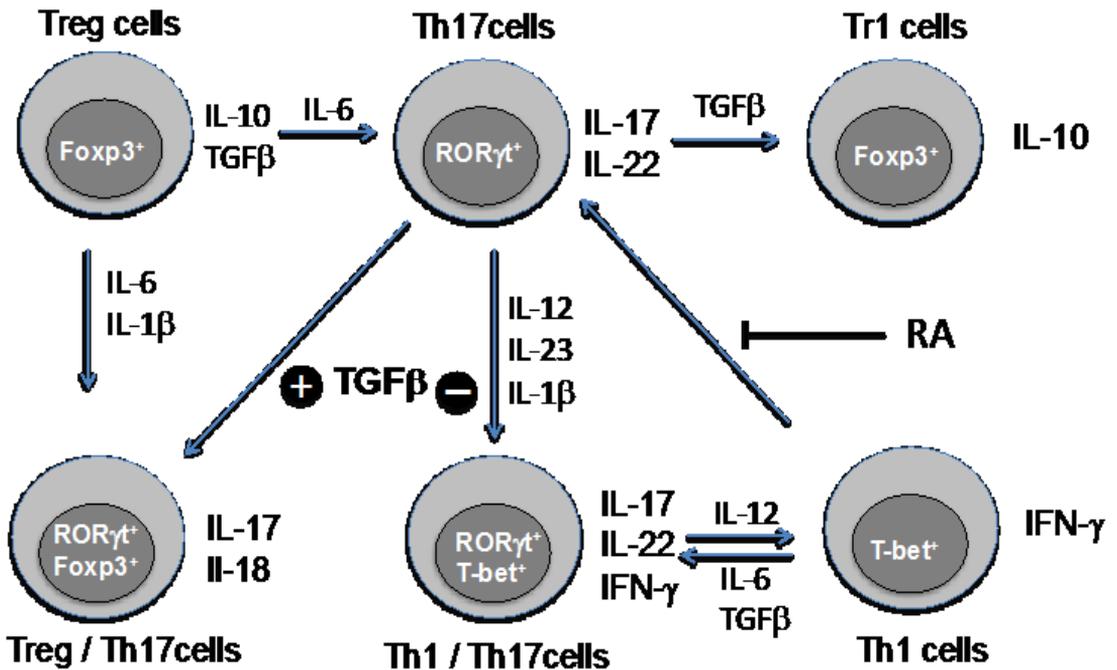


Figure 2: Duality of Th17 cells.

Th17 cells lose stability in the absence of TGFβ and presence of IL-12, IL-23, and IL-1β, favoring IFNγ expression and differentiation into Th1/Th17 cells that produce both Th1 (producing IFNγ) and Th17 (producing IL-17 and IL-22) cytokines. Further augmentation of IL-12 can fully convert Th1/Th17 cells into Th1 cells, whereas this process can be reverted by either TGFβ and IL-6 or in the absence of retinoic acid (RA) in favor of Th1/Th17 or Th17 cells, respectively.

Alternatively, the abundance of TGFβ in the absence of IL-6 drives Th17 cells toward regulatory phenotypes, such as either RORγ^{t+}Foxp3⁺ Treg/Th17 cells or Foxp3⁻ Tr1 cells. If proinflammatory cytokines are present, including either IL-6 or IL-1β and IL-6, Foxp3⁺ Treg have the ability to transdifferentiate into either Th17 or Treg/Th17 cells, respectively.

TRANSDIFFERENTIATION OF TH17 CELLS

Inflammation is a beneficial host response to infection but can contribute to inflammatory disease if unregulated. Th17 cells have been involved in several human inflammatory diseases [24-25]. These cells exhibit both instability and plasticity upon *in vitro* re-stimulation. However, technical limitations have prevented the transcriptional profiling of pre- and post-conversion Th17 cells *ex vivo* during immune responses. Thus, it is unknown whether Th17 cell plasticity merely reflects change in cytokines expression, or if Th17 cells physiologically undergo global genetic reprogramming driving their conversion from one T helper cell type to another, a process known as transdifferentiation [4;26]. Furthermore, although Th17 cell instability/plasticity has been associated with pathogenicity [27], it is unknown whether this could represent a therapeutic

opportunity whereby formerly pathogenic Th17 cells could adopt an anti-inflammatory fate. The transdifferentiation of Th17 into Treg cells was illustrated by a change in their transcriptional profile signature and the acquisition of potent suppressive capacity. Comparisons of the transcriptional profiles of pre- and post-conversion Th17 cells also revealed a role for canonical TGF- β signalling and consequently for the AhR. Thus, Th17 cells transdifferentiate into Treg cells, and may contribute to the resolution of inflammation.

REGULATORY TH17 CELLS?

The relationship between Th17 cells and Treg cells is now the subject of intense research efforts in autoimmune and inflammatory diseases. During polarization, TGF- β and IL-6 polarize naive T cells toward a Th17 phenotype, whereas TGF- β alone induces Treg cells. Thus, it seems that these two 'lineages' share an intermediate stage in their development (Figure 2). Interestingly, Th17 cells that have been matured with TGF- β and IL-6 express strongly genes encoding ROR γ t, IL-17A, IL-17F and IL-21, produce IL-10 and are capable of regulatory functions. The discovery of human IL-17-producing ROR γ t⁺ Foxp3⁺T cells that retain their ability to suppress effector lymphocytes further, supports this 'dual-natured' hypothesis [28-29] (Figure 2).

INCREASED IL-17 LEVEL AND TH17 CELLS IN BD

Multiple independent studies have demonstrated that the level of IL-17A is higher in patients with activeBD [30- 34] compared to healthy controls (P < 0.0001 and P < 0.005). Moreover, the frequency of circulating Th17cells, IL-17A concentration in supernatant of lymphocytes culture, and IL-17A mRNA expression in activated peripheral blood mononuclear cells (PBMCs) are significantly increased (P < 0.0001) [9-11].

Most studies have shown that either the percentage of Th17 cells or the concentration of serum IL-17A correlate positively with BD activity [35-41].

Polarized BD Th17 cells expressed large amounts of transcription factor ROR γ t. In contrast, *in vitro*-treated infliximab Th17 cells expressed less ROR γ t. Stimulation of PBMCs with anti-CD3 and anti-CD28 antibodies resulted in the production of higher IL-17 levels in the cell culture supernatants of PBMCs from BD patients than those obtained from controls (P < 0.001) [36]. Addition of recombinant human interferon- α (rhIFN- α) to this cell culture model decreased significantly IL-17 production while increasing IL-10 production ("both P < 0.001) [36], inducing consequently TGF- β release and Treg cells differentiation. IFN- α activity was mediated via signal transducers and activators of transcription 2 (STAT2) phosphorylation. IFN- α modulated pro- and anti-inflammatory cytokines secreted by T cells increasing IL-10/IL-6 ratio in BD. IFN- α increased IL-10 secretion in each memory (m) subset mTh1, mTh2 and mTh17 [31]. The mechanism by which IFN- α exerts its inhibitory effect on IL-17 was partially mediated by IL-10 and was associated with an up-regulation of the level of p-STAT2.

Recent data demonstrated that increased activation of the Notch pathway is associated with an increased Th17 response in active BD patients [41]. Blocking the Notch pathway can preferentially attenuate the Th17 response by modulating STAT3 phosphorylation [41]. Additionally, the authors showed that decreased expression of miR-23b might contribute to activation of the Notch pathway and the expansion of Th17 cells in BD patients. These interesting results suggested that increased activation of the Notch pathway due to decreased expression of miR-23b might contribute to the pathogenesis of BD [41]. In BD, STAT3 expression was significantly elevated [42] and JAK1/STAT3 signalling pathway was activated, possibly through the activation of Th1/Th17-type cytokines such as IL-2, IFN- γ , IL-6, IL-17 and IL-23 [42].

Increased serum levels of IL-17A were observed in active BD and might serve as markers of disease activity [36; 40]. Moreover, cerebrospinal fluid (CSF) levels of IL-17A were demonstrated to be higher in BD patients with central nervous system involvement compared to NIND (non-inflammatory neurological disease) and HaBD (headache attributed to BD) patients ($P < 0.001$; $P = 0.0021$ respectively) [9]. The relative expression of transcription factors of ROR γ t/FOXP3 ratio (Th17/Treg cells) in CSF was increased in neuro-BD (NBD) patients. These observations of increased expression of IL-17 in blood and in inflammatory sites (CSF) support a Th17-mediated pathogenesis [9]. This suggests the possibility that IL-17A somehow drives central nervous system inflammation in BD [9].

Although Th17 cells are named after their ability to produce IL-17 and represent a main source of IL-17 production, other cells can secrete IL-17. TCR γ δ T cells are an important source of IL-17 during infections [43]. Mice that lack TCR γ δ T cells produce less IL-17. TCR γ δ T cells are activated in BD producing high levels of cytokines [44].

Neutrophils, when stimulated with IL-15, are also able to secrete IL17. Studies have shown that certain memory CD8+ T cells, after stimulation with PMA and ionomycin, can produce IL-17 [45]. BD as systemic inflammatory disorder is characterized by recurrent episodes of acute inflammation consisting mainly of neutrophil infiltration around blood vessels in affected tissues.

Stimulated Natural killer T cells (NKT) IL-23R+ cells, when cultured with IL-23 and anti-CD3, also produce IL-17 [46]. NKT cells that are possibly down regulated by type-I interferon, were also reported in BD [47].

Taken together, these data suggest that IL-17 is not produced solely by a specific set of T cells but a more generic cytokine that is secreted by several cell populations such as TCR γ δ cells, neutrophils and NKT cells in an environment of pro-inflammatory cytokines, leading in turn to propagation of the inflammatory response. In patients with BD, IL-17A promotes the inflammatory process by inducing local production of cytokines and chemokines from multiple cell types including epithelial cells [23; 47- 55]. Geri provided the first evidence of the critical role of IL-21 in driving inflammatory lesions in BD by promoting Th17 effectors and suppressing Treg cells [52].

In BD as reported in autoimmune and inflammatory diseases, Th17 cell duality/plasticity has been associated with pathogenicity and it is unknown whether this could represent a therapeutic opportunity, whereby formerly pathogenic Th17 cells could adopt an anti-inflammatory fate.

ASSOCIATIONS OF GENETIC VARIATIONS OF IL17 WITH BEHÇET'S DISEASE

Behçet's disease is generally considered to be a multifactorial disease with important genetic and environmental components. Genetic polymorphisms of inflammatory cytokines have been associated with BD susceptibility. Recent data suggested that Th17 cell-related genes could act as susceptibility genes for BD in Korean population [53;56], particularly in BD patients with uveitis. Uveitis is a sight-threatening intraocular inflammatory disease that is estimated to account for ~10% of blindness. Based on its etiology, uveitis can be classified into two categories: uveitis related to infection and uveitis that is not. Noninfectious uveitis is frequently associated with autoimmune diseases, including BD, Vogt-Koyanagi-Harada (VKH) syndrome, systemic lupus erythematosus, sarcoidosis, autoimmune hepatitis, and multiple sclerosis [57]. Hou et al. reported that IL17F gene expression was increased in male BD patients [58]. Moreover, this study showed that high copies of IL17F were positively related with the expression of IL17F and may enhance peripheral blood cells proliferation. Such findings were consistent with data shown in SLE patients [59]. IL17 secretion was elevated in uveitis BD and VKH patients [60] supporting the important role of Th17 cells in the pathogenesis of intraocular inflammation.

Considering the role of Th17 cells in experimental autoimmune uveo-retinitis [61], the results suggested that IL17F copy number variant (CNV) might be involved in uveitis via the upregulated expression of IL17F [58]. However, Shu, et al. found no association between BD disease and two single nucleotide polymorphisms (SNPs) of IL-17A and IL-17F whereas a positively association was observed with VKH syndrome [62]. Recent clinical trials of secukinumab, a fully human anti-IL-17A monoclonal antibody, demonstrated its efficacy and safety for the treatment of chronic and active noninfectious uveitis requiring corticosteroid-sparing immunosuppressive therapy [63].

Genome-wide association (GWAS) and replication studies identified in BD a susceptibility locus around STAT4 which expression could regulate IL-17 production [64]. Increased expression of STAT4 was observed in individuals carrying the rs897200 risk genotype AA together with increased IL17 messenger RNA and protein levels. Furthermore, Kim et al [65] demonstrated that interactions of particular IL17A, IL23R, and STAT4 SNPs modulate susceptibility to intestinal BD in the Korean population.

Additional studies are needed to ascertain whether the results presented here can be extrapolated to other ethnic groups in the world.

B AND T LYMPHOCYTE ATTENUATOR IN BD MAY TRIGGER ABNORMAL TH17 IMMUNE RESPONSES

B and T-lymphocyte attenuator (BTLA, also known as CD272) is a member of the B7/CD28 superfamily and was first identified as an inhibitory receptor on T cells on the basis of the enhanced T cell responses that were observed in Btla-knockout mice [66]. Its ligand herpes virus entry mediator (HVEM, also known as TNFRSR14) is a member of TNF/TNFR superfamily. BTLA is broadly expressed on B cells, T cells, DCs, macrophages, and NK cells [67].

Recent studies have investigated whether BTLA activation could be exploited to inhibit the development of abnormal immune responses in BD patients [68]. BTLA mRNA and protein expression were found significantly decreased in BD patients with active ocular inflammation compared to the normal controls ($P < 0.001$). Decreased percentages of BTLA^{high} cells and CD4+BTLA^{high} cells were also observed in peripheral blood of these patients ($P = 0.011$; $P = 0.010$) while IL-17-positive cells were significantly increased in the CD4+BTLA^{lo} cells population ($P < 0.01$, $P < 0.001$) [68]. In BD patients and controls, the agonistic anti-BTLA antibody reduced the frequency of IL-17- and IFN- γ -producing CD4+ T cells ($P < 0.05$, $P < 0.05$) and inhibited the production IL-17 cytokine ($P < 0.05$) ($P < 0.05$). BTLA-mediated inhibition of T cell activation occurred during both primary CD4+ T cell responses and secondary CD4+ and CD8+ T cell responses, suggesting that BTLA ligation sends a constitutive “off” signal to T cells and thus might play an important role in the maintenance of T cell tolerance [69].

FACTORS SUPPRESSING TH17 PRODUCTION IN BD

Vitamin D (Vit D) deficiency was observed in BD patients and correlated with inflammatory status [70] while its supplementation had favourable effects on endothelial function [71]. Single nucleotide polymorphisms (SNPs) of Vit D family genes increased the susceptibility risk for BD [72-73]. Recent data showed that vitamin D was an important promoter of T cell regulation *in vivo* in BD patients and modulated inflammatory mediators production. Its deficiency was associated with increased levels of Th17 cell cytokines (IL-17 and IL-21) [70]. IL-17+ cells were negatively correlated with vitamin D levels ($r = -0.462$; $P = 0.0403$). The inhibitory effect of Vit D on the Th17 and Th1 responses was mediated via both T cells and DCs while IFN regulatory factor 8 (IRF-8) pathway was involved in inhibition of Th17 cell differentiation [74].

Certain mediators exert broad inhibitory properties on the innate inflammatory and acquired immune responses. Studies were set up to investigate the expression of IL-27 and IL-37 in Behçet disease (BD) and to explore their possible regulatory role during inflammation, particularly on the suppression of Th17 cells activity.

IL-27, a heterodimeric cytokine composed of two subunits: p28 (IL-27p28) and the Epstein-Barr virus-induced gene 3 (EBI3), is produced mainly by activated antigen-presenting cells (APCs) [75]. The IL-27 receptor is widely expressed on naïve T cells, natural killer cells, mast cells, monocytes, keratinocytes, vascular endothelium, activated B cells, DCs and Langerhans cells. Low

IL-27 expression was reported in BD patients compared to inactive BD patients ($P = 0.009$) and to healthy controls ($P < 0.001$) [76]. Recombinant IL-27 was able to inhibit Th17 differentiation in both BD patients and healthy controls. Results from Wang [76] provided evidence that the negative regulatory effect of IL-27 on Th1 and Th17 cells was mediated via DCs and suggested the involvement of the IRF-8 pathway in this suppressive effect. A decreased expression of IL-27 was associated with active intraocular inflammation in BD [76].

IL-37 that belongs to the members of the IL-1 family has been described as an anti-inflammatory cytokine in several autoimmune/Inflammatory diseases. IL-37 can be induced in various types of cells such as PBCs, epithelial cells, DCs, monocytes and keratinocytes [77]. IL-37 was significantly decreased in active BD patients compared to healthy controls ($P < 0.0001$), and could effectively decrease the productions of pro-inflammatory cytokines TNF- α , IL-6, IL-1 β and IL-17. Furthermore, recombinant IL-37 (rIL-37) exerted a more suppressive effect on IL-17 production in active BD patients than in healthy controls [78]. Ye et al. reported that rIL-37-treated DCs in BD notably inhibited Th17 and Th1 cell responses as compared to control DCs [79]. These findings suggest that IL-37 could play a potent immunosuppressive role in the pathogenesis of BD via the down regulation of IL-17.

In recent trials, anti-IL-17 and anti-IL-17 receptor antibodies induced a rapid and robust clinical improvement in several autoimmune diseases such as rheumatoid arthritis, ankylosing spondylitis, psoriasis [80-82]. These antibodies were not tried in BD. Interferon alpha (IFN- α) is an effective treatment for patients with active Behçet disease (BD). Besides its antiviral property, IFN- α can generate an anti-inflammatory environment or inhibit specific inflammatory T cells such as Th1 and Th17 cells [34].

Anti-TNF- α therapy inhibited *in vivo* local IL-17 production in ocular fluids from BD patients with uveitis. Moreover, *in vitro* infliximab reduced RORyt expression and IL-17 production from BD TH17 cells [38].

DISTURBED BALANCE BETWEEN TREG AND TH17 CELLS IN BD

The pathologic hallmarks of BD are based on a dysregulated immune response, and include subsequent inflammatory tissue injury. It is widely acknowledged that the level of IL-17A and the percentage of Th17 cells are increased in BD animal models and BD patients. Regulatory T cells as well as Th17 cells have been shown to possess a certain degree of plasticity (Figure 3). It has been described that mouse Foxp3+ Treg cells are able to transdifferentiate into Th17-like cells due to the action of IL-6 in the absence of TGF β [83-84]. In humans, Treg cells are able to adapt a Th17-like phenotype, which is accompanied with the production of IL-17 [8]. It appears that Treg cells that produce IL-17 can retain their suppressive function until they are triggered by IL-6 and IL-1 β [19]. Dynamic changes in the cytokine milieu may transiently disturb the balance between Th17 cells and Treg cells, thereby driving flares of active BD disease [52,85,86].

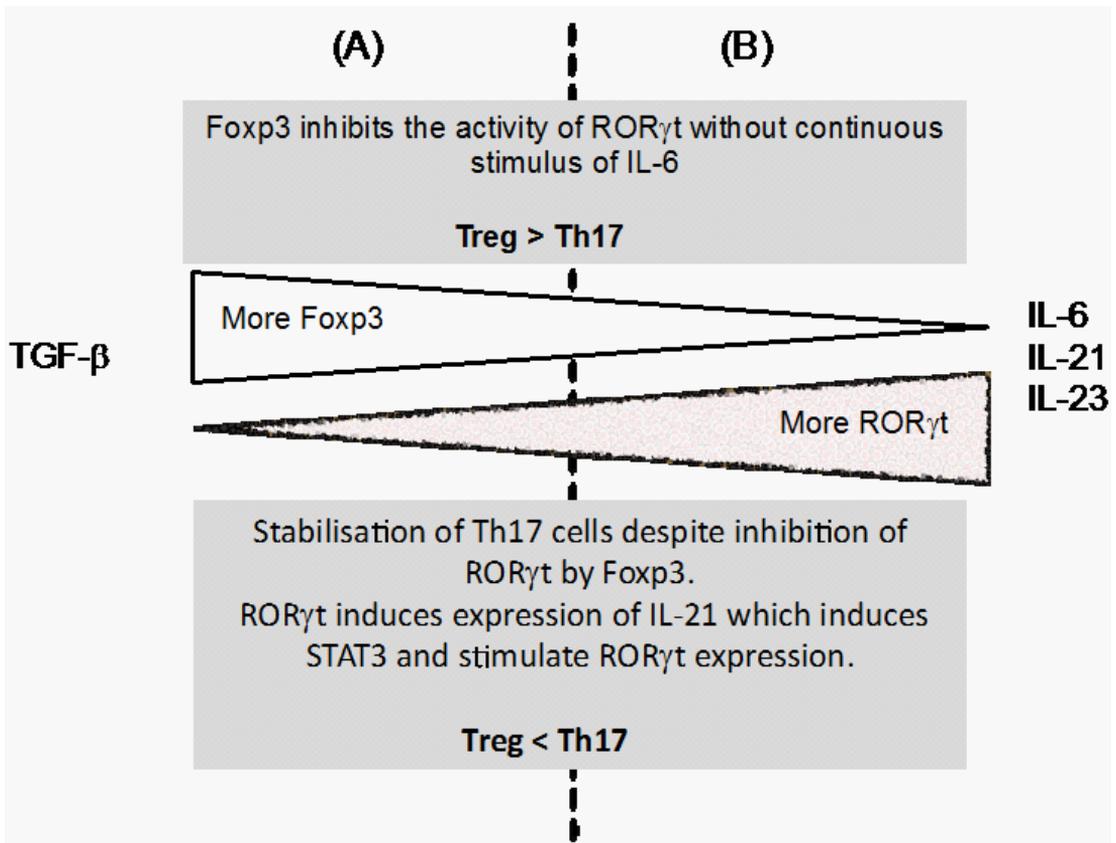


Figure 3: Hypothetical Plasticity between Treg and Th17 cells in Behçet disease.

(A): Possible transcription factor interactions regulating intermediate CD4+ T cell transitions. Foxp3 inhibits the activity of ROR γ t, without a reciprocal inhibition or feedback from ROR γ t. This circuit might favor Treg over Th17 development, making the Treg state more stable. Without continuous stimulus by IL-6 to induce ROR γ t, Foxp3 might tend to repress Th17 development. (B) Second possibility, Th17 state is stabilized despite inhibition of ROR γ t by Foxp3 owing to a known feedback loop, in which ROR γ t induces expression of IL-21, which acts in an autocrine manner to further induce STAT3 and stimulate ROR γ t expression. This feedback stabilizes the Th17 state relative to the Treg state.

CONCLUDING REMARKS

The discovery of Th17 cells has led to a plethora of studies targeting their role in BD inflammatory pathways. A number of basic findings have also lightened the autoimmune / inflammatory field through characterization of Th17 cells as a distinct subset that builds on the Th1/Th2 paradigm. However, as discussed herein, the role of the expanded Th17 subpopulation in BD immunity remains ambiguous and appears to be dependent upon several factors such as co-stimulatory molecules, signal transducers and activators of transcription (STATs).

Plasticity between Treg and Th17 likely occurs in the context of dynamic changes in the inflammatory environment. Thus, pro-inflammatory stimuli may promote conversion of immune-suppressive regulatory T cells into pro-inflammatory Th17 cells, while resolution of inflammation may trigger or even require the alternate shift from Th17 to Treg. This concept is just becoming appreciated and requires further study to correlate both causes and outcomes.

A number of crucial questions remain to be answered. How might Th17/Treg imbalance be induced and lead to disease exacerbation, inducing immune pathogenesis of uveitis, central nervous system involvement and lung manifestations? Further understanding of the mechanisms of Th17/Treg-mediated inflammatory immune responses, in tilting the balance between destructive inflammation and homeostasis, may open new lines of investigation for BD treatment in the future.

References

1. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL, et al. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J Immunol.* 2005; 175: 5-14.
2. Annunziato F and Romagnani S. Do studies in humans better depict Th17 cells? *Blood.* 2009; 114: 2213-2219.
3. Kullberg MC, Jankovic D, Feng CG, Hue S, Gorelick PL, et al. IL-23 plays a key role in Helicobacter hepaticus-induced T cell-dependent colitis. *J Exp Med.* 2006; 203: 2485-2494.
4. Graf T and Enver T. Forcing cells to change lineages. *Nature.* 2009; 462: 587-594.
5. Damsker JM, Hansen AM, Caspi RR. Th1 and Th17 cells: adversaries and collaborators. *Ann N Y Acad Sci.* 2010; 1183: 211-221.
6. Lee YK, Turner H, Maynard CL, Oliver JR, Chen D, et al. Late developmental plasticity in the T helper 17 lineage. *Immunity.* 2009; 30: 92-107.
7. Rebhahn JA, Deng N, Sharma G, Livingstone AM, Huang S, et al. An animated landscape representation of CD4+ T-cell differentiation, variability, and plasticity: insights into the behavior of populations versus cells. *Eur J Immunol.* 2014; 44: 2216-2229.
8. Koenen HJ, Smeets RL, Vink PM, van Rijssen E, Boots AM, et al. Human CD25highFoxp3pos regulatory T cells differentiate into IL-17-producing cells. *Blood.* 2008; 112: 2340-2352.
9. Hamzaoui K, Haghighi AB, Ghorbel IB, Houman H. RORC and Foxp3 axis in cerebrospinal fluid of patients with neuro-Behçet's disease. *J. Neuroimmunol.* 2011; 233: 249-253.
10. Hamzaoui K, Bouali E, Ghorbel I, Khanfir M, Houman H, et al. Expression of Th-17 and RORγt mRNA in Behçet's Disease. *Med. Sci. Monit.* 2011; 17: CR227-234.
11. Hamzaoui K. Th17 cells in Behçet's disease: a new immunoregulatory axis. *Clin Exp Rheumatol.* 2011; 29: S71-76.
12. Ueno A, Ghosh A, Hung D, Li J, Jijon H. Th17 plasticity and its changes associated with inflammatory bowel disease. *World J Gastroenterol.* 2015; 21: 12283-12295.
13. Cosmi L, Santarlasci V, Maggi L, Liotta F, Annunziato F. Th17 plasticity: pathophysiology and treatment of chronic inflammatory disorders. *Curr Opin Pharmacol.* 2014; 17: 12-16.
14. Huber S, Gagliani N, Flavell RA. Life, death, and miracles: TH17 cells in the intestine. *Eur J Immunol.* 2012; 42: 2238-2245.
15. Roncarolo MG, Battaglia M. Regulatory T-cell immunotherapy for tolerance to self antigens and alloantigens in humans. *Nature Rev Immunol.* 2007; 7: 585 -598.
16. Heinemann C, Heink S, Petermann F, Vasanthakumar A, Rothhammer V, et al. IL-27 and IL-12 oppose pro-inflammatory IL-23 in CD4+ T cells by inducing Blimp1. *Nature Commun.* 2014; 5: 3770.
17. Okamura T, Sumitomo S, Morita K, Iwasaki Y, Inoue M, et al. TGF-β3-expressing CD4+CD25-LAG3+ regulatory T cells control humoral immune responses. *Nature Commun.* 2015; 6: 6329.
18. Littman DR, Rudensky AY. TH17 and regulatory T cells in mediating and restraining inflammation. *Cell.* 2010; 140: 845-858.
19. Zhou L, Lopes JE, Chong MM, Ivanov II, Min R, et al. TGF-beta-induced Foxp3 inhibits Th17 cell differentiation by antagonizing RORgammat function. *Nature.* 2008; 453: 236-240.

20. Beriou G, Costantino CM, Ashley CW, Yang L, Kuchroo VK, et al. IL-17-producing human peripheral regulatory T cells retain suppressive function. *Blood*. 2009; 113: 4240 - 4249.
21. Esplugues E, Huber S, Gagliani N, Hauser AE, Town T, et al. Control of TH17 cells occurs in the small intestine. *Nature*. 2011; 475: 514–518.
22. Zielinski CE, Mele F, Aschenbrenner D, Jarrossay D, Ronchi F, et al. Pathogen-induced human TH17 cells produce IFN- γ or IL-10 and are regulated by IL-1 β . *Nature*. 2012; 484: 514-518.
23. Burkett PR, Meyer zuHorste G, Kuchroo VK. Pouring fuel on the fire: Th17 cells, the environment, and autoimmunity. *J Clin Invest*. 2015; 125: 2211-2219.
24. Annunziato F, Cosmi L, Santarlasci V, Maggi L, Liotta F, et al. Phenotypic and functional features of human Th 17 cells. *J Exp Med*. 2007; 204: 1849-1861.
25. Hirota K, Duarte JH, Veldhoen M, Hornsby E, Li Y, et al. Fate mapping of IL-17-producing T cells in inflammatory responses. *Nature Immunol*. 2011; 12: 255-263.
26. Komatsu N, Mariotti-Ferrandiz ME, Wang Y, Malissen B, Waldmann H, et al. Heterogeneity of natural Foxp3+ T cells: a committed regulatory T-cell lineage and an uncommitted minor population retaining plasticity. *Proc Natl Acad Sci USA*. 2009; 106: 1903-1908.
27. Gagliani N, Vesely AMC, Iseppon A, Brockmann L, Xu H, et al. Th17 cells transdifferentiate into regulatory T cells during resolution of inflammation. *Nature*. 2015; 523: 221-225.
28. Lubberts E. The IL-23-IL-17 axis in inflammatory arthritis. *Nat Rev Rheumatol*. 2015; 11: 415-429.
29. Takeuchi M, Kastner DL, Remmers EF. The immunogenetics of Behçet's disease: A comprehensive review. *J Autoimmun*. 2015; 64: 137-148.
30. Direskeneli H, Fujita H, Akdis CA. Regulation of TH17 and regulatory T cells in patients with Behçet disease. *J Allergy Clin Immunol*. 2011; 128: 665-666.
31. Na SY, Park MJ, Park S, Lee ES. Up-regulation of Th17 and related cytokines in Behçet's disease corresponding to disease activity. *Clin Exp Rheumatol*. 2013; 31: 32-40.
32. Ozyurt K, Celik A, Sayarlioglu M, Colgecen E, Inci R, et al. Serum Th1, Th2 and Th17 cytokine profiles and alpha-enolase levels in recurrent aphthous stomatitis. *J Oral Pathol Med*. 2014; 43: 691-695.
33. Cetin AE, Cosan F, Ceffe A, Deniz G. IL-22-secreting Th22 and IFN- γ secreting Th17 cells in Behçet's disease. *Mod Rheumatol*. 2014; 24: 802-807.
34. Touzot M, Cacoub P, Bodaghi B, Soumelis V, Saadoun D. IFN- α induces IL-10 production and tilt the balance between Th1 and Th17 in Behçet disease. *Autoimmun Rev*. 2015; 14: 370-375.
35. Hamzaoui K, Hamzaoui A, Guemira F, Bessioud M, Hamza M, et al. Cytokine profile in Behcet's disease patients. Relationship with disease activity. *Scand J Rheumatol*. 2002; 31: 205–210.
36. Liu X, Yang P, Wang C, Li F, Kijlstra A. IFN-alpha blocks IL-17 production by peripheral blood mononuclear cells in Behcet's disease. *Rheumatology (Oxford)*. 2011; 50: 293-298.
37. Shimizu J, Izumi T, Arimitsu N, Fujiwara N, Ueda Y, et al. Skewed TGF β /Smad signalling pathway in T cells in patients with Behçet's disease. *Clin Exp Rheumatol*. 2012; 30: S35-39.
38. Sugita S, Kawazoe Y, Imai A, Yamada Y, Horie S, et al. Inhibition of Th17 differentiation by anti-TNF-alpha therapy in uveitis patients with Behçet's disease. *Arthritis Res Ther*. 2012; 14: R99.
39. Dagur PK, Biancotto A, Stansky E, Sen HN, Nussenblatt RB, et al. Secretion of interleukin-17 by CD8+ T cells expressing CD146 (MCAM). *Clin Immunol*. 2014; 152: 36-47.
40. Mesquida, M, Molins B, Llorenç V, Hernández MV, Espinosa G, et al. Current and future treatments for Behçet's uveitis: road to remission. *Int Ophthalmol*. 2014; 34: 365-381.
41. Qi J, Yang Y, Hou S, Qiao Y, Wang Q, Yu H, et al. Increased Notch pathway activation in Behçet's disease. *Rheumatology (Oxford)*. 2014; 53: 810-820.
42. Tulunay A, Dozmorov MG, Ture-Ozdemir F, Yilmaz V, Eksioğlu-Demiralp E, et al. Activation of the JAK/STAT pathway in Behcet's disease. *Genes Immun*. 2015; 16: 170-175.
43. Pineton de Chambrun M, Wechsler B, Geri G, Cacoub P, et al. New insights into the pathogenesis of Behçet's disease. *Autoimmun Rev*. 2012; 11: 687-698.
44. Ferretti S, Bonneau O, Dubois GR, Jones CE, Trifilieff A. IL-17, produced by lymphocytes and neutrophils, is necessary for lipopolysaccharide-induced airway neutrophilia: IL-15 as a possible trigger. *J Immunol*. 2003; 170: 2106-2112.

45. Neves FS and Spiller F. Possible mechanisms of neutrophil activation in Behçet's disease. *Int Immunopharmacol.* 2013; 17: 1206-1210.
46. Yilmaz S, Cinar M, Pekel A, Simsek I, Musabak U, et al. The expression of transmembrane and soluble CXCL16 and the relation with interferon-alpha secretion in patients with Behçet's disease. *Clin Exp Rheumatol.* 2013; 31: 84-87.
47. Rachitskaya AV, Hansen AM, Horai R, Li Z, Villasmil R, et al. Cutting edge: NKT Cells Constitutively Express IL-23 Receptor and ROR γ t and Rapidly Produce IL-17 upon Receptor Ligation in an IL-6-Independent Fashion. *J Immunol.* 2008; 180: 5167-5171.
48. Atzeni F, Boiardi L, Casali B, Farnetti E, Nicoli D, et al. CC chemokine receptor 5 polymorphism in Italian patients with Behçet's disease. *Rheumatology (Oxford).* 2012; 51: 2141-2145.
49. Ferran M, AB Galván, A Giménez-Arnau, RM Pujol, et al. Production of Interleukin-8 by Circulating CLA+ T Cells with Skin Tropism in Patients with Psoriasis and in Healthy Controls. *Actas Dermosifiliogr.* 2010; 101: 151-155.
50. Hou S, Yang P, Du L, Jiang Z, Mao L, et al. Monocyte chemoattractant protein-1 -2518 A/G single nucleotide polymorphism in Chinese Han patients with ocular Behçet's disease. *Hum Immunol.* 2010; 71: 79-82.
51. El-Asrar AM, Al-Obeidan SS, Kangave D, Geboes K, Opdenakker G, et al. CXC chemokine expression profiles in aqueous humor of patients with different clinical entities of endogenous uveitis. *Immunobiology.* 2011; 216: 1004-1009.
52. Geri G, Terrier B, Rosenzweig M, Wechsler B, Touzot M, et al. Critical role of IL-21 in modulating TH17 and regulatory T cells in Behçet disease. *J Allergy Clin Immunol.* 2011; 128: 655-664.
53. Kim ES, Kim SW, Moon CM, Park JJ, Kim TI, et al. Interactions between IL17A, IL23R, and STAT4 polymorphisms confer susceptibility to intestinal Behçet's disease in Korean population. *Life Sci.* 2012a; 90: 740-746.
54. Zhou ZY, Chen SL, Shen N, Lu Y. Cytokines and Behçet's disease. *Autoimmun Rev.* 2012; 11: 699-704.
55. Hamzaoui K, Borhani-Haghighi A, Kaabachi W, Hamzaoui A. Increased interleukin 33 in patients with neuro-Behçet's disease: correlation with MCP-1 and IP-10 chemokines. *Cell Mol Immunol.* 2014; 11: 613-616.
56. Jang WC, Nam YH, Ahn YC, Lee SH, Park SH, et al. Interleukin-17F gene polymorphisms in Korean patients with Behçet's disease. *Rheumatol Int.* 2008; 29: 173-178.
57. Barisani-Asenbauer T, Maca SM, Mejdoubi L, Emminger W, Machold K, et al. Uveitis- a rare disease often associated with systemic diseases and infections- a systematic review of 2619 patients. *Orphanet J. Rare Dis.* 2012; 7: 57-63.
58. Hou S, Liao D, Zhang J, Fang J, Chen L, et al. Genetic variations of IL17F and IL23A show associations with Behçet's disease and Vogt-Koyanagi-Harada syndrome. *Ophthalmology.* 2015; 122: 518-523.
59. Yu B, Guan M, Peng Y, Shao Y, Zhang C, et al. Copy number variations of interleukin-17F, interleukin-21 and interleukin-22 are associated with systemic lupus erythematosus. *Arthritis Rheum.* 2011; 63: 3487-3492.
60. Chi W, Zhu X, Yang P, Liu X, Zhou H, et al. Upregulated IL-23 and IL-17 in Behçet patients with active uveitis. *Invest Ophthalmol Vis Sci.* 2008; 49: 3058-3064.
61. Amadi-Obi A, Yu CR, Liu X, Mahdi RM, Clarke GL, et al. TH17 cells contribute to uveitis and scleritis and are expanded by IL-2 and inhibited by IL-27/STAT1. *Nat Med.* 2007; 13: 711-718.
62. Shu Q, Yang P, Hou S, Li F, Chen Y, et al. Interleukin-17 gene polymorphism is associated with Vogt-Koyanagi-Harada syndrome but not with Behçet's disease in a Chinese Han population. *Hum Immunol.* 2010; 71: 988-991.
63. Letko E, Yeh S, Foster CS, Pleyer U, Brigell M, et al. Efficacy and safety of intravenous secukinumab in noninfectious uveitis requiring steroid-sparing immunosuppressive therapy. *Ophthalmology.* 2015; 122: 939-948.
64. Hou S, Yang Z, Du L, Jiang Z, Shu Q, et al. Identification of a susceptibility locus in STAT4 for Behçet's disease in Han Chinese in a genome-wide association study. *Arthritis Rheum.* 2012; 64: 4104-4113.
65. Kim SK, Jang WC, Ahn YC, Lee SH, Lee SS, et al. Promoter -2518 single nucleotide polymorphism of monocyte chemoattractant protein-1 is associated with clinical severity in Behçet's disease. *Inflamm Res.* 2012; 61: 541-545.
66. Watanabe N, Gavrieli M, Sedy JR, Yang J, Fallarino F, et al. BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. *Nat Immunol.* 2003; 4: 670-679.
67. Hurchla MA, Sedy JR, Gavrieli M, Drake CG, Murphy TL, et al. B and T lymphocyte attenuator exhibits structural and expression polymorphisms and is highly Induced in anergic CD4+ T cells. *J. Immunol.* 2005; 174: 3377-3385.
68. Ye Z, Deng B, Wang C, Zhang D, Kijlstra A, et al. Decreased B and T lymphocyte attenuator in Behçet's disease may trigger abnormal Th17 and Th1 immune responses. *Sci Rep.* 2016; 6: 20401.
69. Otsuki N, Kamimura Y, Hashiguchi M, Azuma M. Expression and function of the B and T lymphocyte attenuator (BTLA/CD272) on human T cells. *Biochem Biophys Res Commun.* 2006; 344: 1121-1127.

70. Hamzaoui K, Dhifallah IB, Karray E, Sassi FH, Hamzaoui A. Vitamin D modulates peripheral immunity in patients with Behçet's disease. *Clin Exp Rheumatol*. 2010; 28: S50-57.
71. Can M, Gunes M, Haliloglu OA, Haklar G, Inanç N. Effect of vitamin D deficiency and replacement on endothelial functions in Behçet's disease. *Clin Exp Rheumatol*. 2012; 30: S57-61.
72. Fang J, Hou S, Xiang Q, Qi J, Yu H, et al. Polymorphisms in genetics of vitamin D metabolism confer susceptibility to ocular Behçet disease in a Chinese Han population. *Am J Ophthalmol*. 2014; 157: 488-494.e6.
73. Tizaoui K, Kaabachi W, Salah OM, Amor BA, Hamzaoui A, et al. Vitamin D receptor TaqI and Apal polymorphisms: a comparative study in patients with Behçet's disease and Rheumatoid arthritis in Tunisian population. *Cell Immunol*. 2014; 290: 66-71.
74. Tian Y, Wang C, Ye Z, Xiao X, Kijlstra A, et al. Effect of 1, 25-dihydroxyvitamin D3 on Th17 and Th1 response in patients with Behçet's disease. *Invest Ophthalmol Vis Sci*. 2012; 53: 6434-6441.
75. Pflanz S, Timans JC, Cheung J, Rosales R, Kanzler H, et al. IL-27, a heterodimeric cytokine composed of EBI3 and p28 protein, induces proliferation of naive CD4+ T cells. *Immunity*. 2002; 16: 779-790.
76. Wang C, Tian Y, Ye Z, Kijlstra A, Zhou Y, et al. Decreased interleukin 27 expression is associated with active uveitis in Behçet's disease. *Arthritis Res Ther*. 2014; 16: R117.
77. Nold MF, Nold-Petry CA, Zepp JA, Palmer BE, Buffer P, et al. IL-37 is a fundamental inhibitor of innate immunity *Nat. Immunol*. 2010; 11: 1014-1022.
78. Bouali E, Kaabachi W, Hamzaoui A, Hamzaoui K. Interleukin-37 expression is decreased in Behçet's disease and is associated with inflammation. *Immunol Lett*. 2015; 167: 87-94.
79. Ye Z, Wang C, Kijlstra A, Zhou X, Yang P. A possible role for interleukin 37 in the pathogenesis of Behçet's disease. *Curr Mol Med*. 2014; 14: 535-542.
80. Spuls PI and Hooft L. Brodalumab and ixekizumab, antiinterleukin-17-receptor antibodies for psoriasis: a critical appraisal. *Br. J. Dermatol*. 2012; 167: 710-713.
81. Genovese MC, Durez P, Richards HB, Supronik J, Dokoupilova E, et al. Efficacy and safety of secukinumab in patients with rheumatoid arthritis: a phase II, dose-finding, double-blind, randomised, placebo controlled study. *Ann Rheum Dis*. 2013; 72: 863-869.
82. Baeten D, Baraliakos X, Braun J, Sieper J, Emery P, et al. Anti-interleukin-17A monoclonal antibody secukinumab in treatment of ankylosing spondylitis: a randomised, double-blind, placebo-controlled trial. *Lancet*. 2013; 382: 1705-1713.
83. Xu L, Kitani A, Fuss I, Strober W. Cutting edge: regulatory T cells induce CD4+CD25-Foxp3- T cells or are self-induced to become Th17 cells in the absence of exogenous TGF-beta. *J Immunol*. 2007; 178: 6725-6729.
84. Yang XO, Nurieva R, Martinez GJ, Kang HS, Chung Y, et al. Molecular antagonism and plasticity of regulatory and inflammatory T cell programs. *Immunity*. 2008; 29: 44 -56.
85. Hamzaoui K and Hamzaoui A. Immunological responses in patients with Behçet's disease: advances in understanding. *Expert Rev Ophthalmol*. 2012; 7: 261-270.
86. Barry RJ, Alsalem JA, Faassen J, Murray PI, Curnow SJ, et al. Association analysis of TGFBR3 gene with Behçet's disease and idiopathic intermediate uveitis in a Caucasian population. *Br J Ophthalmol*. 2015; 99: 696-699.